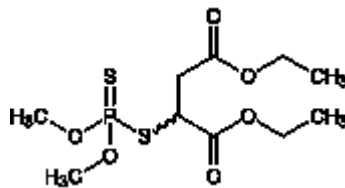




# Fastsættelse af kvalitetskriterier for vandmiljøet

## malathion

CAS nr. 121-75-5



Vandkvalitetskriterium	VKK <sub>ferskvand</sub>	0,003 µg/l
Vandkvalitetskriterium	VKK <sub>saltvand</sub>	0,0006µg/l
Korttidsvandkvalitetskriterium	KVKK <sub>ferskvand</sub>	0,4 µg/l
Korttidsvandkvalitetskriterium	KVKK <sub>saltvand</sub>	0,4 µg/l
Sedimentkvalitetskriterium	SKK <sub>ferskvan</sub>	0,04 µg/kg tørvægt (5% OC)
Sedimentkvalitetskriterium	SKK <sub>ferskvan</sub>	0,8 µg/kg tørvægt x f <sub>oc</sub>
Sedimentkvalitetskriterium	SKK <sub>saltvand</sub>	0,007 µg/kg tørvægt (5% OC)
Sedimentkvalitetskriterium	SKK <sub>saltvand</sub>	0,14 µg/kg tørvægt x f <sub>oc</sub>
Biota-kvalitetskriterium, sekundær forgiftning	BKK <sub>sek.forgiftn.</sub>	0,27 mg/kg musling (bløddele), vådvægt
Biota-kvalitetskriterium, human konsum	HKK	3,7 mg/kg fiskeriprodukter, vådvægt

Januar 2023

*Databladet er i april 2024 opdateret i forhold til at tydeliggøre ved hvilket organisk kulstof (OC) indhold sedimentkvalitetskriterierne er bestemt ved.*

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# Forord

Et kvalitetskriterium i vandmiljøet er det højeste koncentrationsniveau, ved hvilket der skønnes, ikke at forekomme uacceptable negative effekter på vandøkosystemer.

Miljøstyrelsen (MST) udarbejder kvalitetskriterier for kemikalier i vandsøjlen, i sediment, i dyr og planter (biota) og for human konsum.

Miljøstyrelsen bruger kvalitetskriterierne som det faglige grundlag til at kunne fastsætte miljøkvalitetskrav, hvorved der forstås den endelige koncentration af et bestemt forurenende stof i vand, sediment eller biota, som ikke må overskrides af hensyn til beskyttelsen af miljøet og menneskers sundhed.

Metodikken, der anvendes til udarbejdelse af miljøkvalitetskrav er harmoniseret i EU og baserer sig på vandrammedirektivet (EU, 2000), EU's vejledning til fastsættelse af kvalitetskriterier i vandmiljøet (EU, 2018a) og Miljøstyrelsens vejledning til fastsættelse af vandkvalitetskriterier (Miljøstyrelsen, 2004). Metodikken er endvidere i overensstemmelse med EU's vejledning til risikovurdering under REACH forordningen (EU, 2008).

Den sidste litteratursøgning er foretaget oktober 2022.

# English Summary and Conclusions

Derivation of environmental quality standards (EQS) for the aquatic environment is following the EU Guidance Document No. 27. Technical Guidance Document (TGD) for Deriving Environmental Quality Standards (EU, 2018).

Primary reference sources were FMC 2019, and US EPA's database ECOTOX. Data from FMC 2019 were not further checked for reliability as that dataset already has been through such a process.

Especially concerning sediment toxicity, searches have also been made in the following databases: Agris, Biological Abstracts, GeoScience World, PubMed and Google Scholar.

Statistically, differences between sensitivities of freshwater and marine organisms were insignificant, in the case of chronic as well as acute aquatic toxicity. The marine and freshwater datasets may thus be pooled.

## **AA-EQS for water**

For **chronic studies**, all other references were, as far as possible, procured and the reliability assessed (CRED), and only studies with reliability indexes 1 or 2 were accepted.

In ECOTOX the chronic studies were first sorted according to duration of study (should be "long term"). Further, studies where concentrations were not measured were not taken on board unless flow through or renewal was employed. If a formulation or a low purity of the substance was employed, then the study was also not accepted unless concentrations had been measured. Only types of effects that can be related to effects on populations were accepted.

The combined chronic dataset includes 23 species representing 9 major taxonomic groups, and a species sensitivity distribution analysis (SSD) can be performed.

The resulting chronic  $HC_5^{\text{freshwater} + \text{saltwater}} = 0.0141 \mu\text{g/l}$  (90% CI: 0.00063-0.126).

Lognormality is accepted at all significance levels listed in ETx. The standard deviation (s) of  $\log_{10}$  transformed data is 2.04.

The standard deviation is high. Further, the lowest  $EC_{10}$  or NOEC is much lower than the  $HC_5$  (0.0008  $\mu\text{g/l}$ ), and a number of species that are most sensitive in the acute dataset are not represented in the chronic dataset. Therefore, the assessment factor (AF) is not reduced from the default of 5.

Only one species that, according to EU 2018a, can be regarded as representing a specific marine group is included in the chronic dataset. Therefore, an extra AF of 5 is applied for estimation of the marine QS.

**AA-EQS<sub>freshwater</sub> = 0.014  $\mu\text{g/l}$  / 5 = 0.0028  $\mu\text{g/l}$   $\approx$  0.003  $\mu\text{g/l}$**

**AA-EQS<sub>saltwater</sub> = 0.0028  $\mu\text{g/l}$  / 5 = 0.00056  $\mu\text{g/l}$   $\approx$  0.0006  $\mu\text{g/l}$**

### MAC-EQS for water

For **acute studies** the dataset in ECOTOX is huge (1795). The sorting criteria were the same as for chronic studies except that up to two days test without measurements of the substance were accepted because we are dealing with acute (short-term) effects, and the half-life of malathion probably is around 2 days. This means that for many species where the standard is a 4-day test, 2-day tests have been accepted, giving a bias towards greater values. On the other hand, where multiple values occurred for the same species, in ECOTOX, the lowest value was chosen. When multiple values were available from other sources the geometric mean was used – given the studies were performed under the same general conditions and with the same endpoints.

After the first sorting the acute database was reduced to 311 values. Following this, the studies of the lowest and largest values were retrieved and subject to CRED assessment, in all 47 studies covering 58 values.

The combined acute dataset encompasses 272 species representing 16 major taxonomic groups, including groups that are expected to contain some of the most sensitive species i.e., insects and crustaceans. An SSD analysis is thus relevant.

The acute  $HC_5 = 0.88 \mu\text{g/l}$ , with  $s = 1.47$ .

However, the distribution of data is clearly two-peaked, and therefore an SSD based only on data from the lower group was performed. The resulting  $HC_5$  was  $1.2 \mu\text{g/l}$ , and the dataset included data for 159 species and 7 major taxonomic groups. Crustacea and insects were richly represented with a variety of classes and orders, respectively.

Given the large number of species and major taxonomic groups, including the most sensitive the default AF of 10 is reduced to 3.

There are multiple specific marine species in the dataset, so the  $MAC_{\text{freshwater}} = MAC_{\text{saltwater}}$ .

$$MAC-EQS_{\text{freshwater}} = 1.2 \mu\text{g/l} / 3 = 0.4 \mu\text{g/l}$$

$$MAC-EQS_{\text{saltwater}} = 1.2 \mu\text{g/l} / 3 = 0.4 \mu\text{g/l}$$

### QS for sediment

No reliable tests with organisms in sediment were found and the EqP method was employed.

An EU standard sediment with 5% organic carbon (OC) and a  $K_{OC}$  value of 216 l/kg (geometric mean) is used. The partition coefficient between solid and water in sediment,  $K_{p_{\text{sed}}}$  is determined based on the following equation:

$$K_{p_{\text{sed}}} = F_{OC_{\text{sed}}} \times K_{OC} = 0.05 \times 216 \text{ l/kg} = 10.8 \text{ l/kg}$$

The partition coefficient between sediment and water  $K_{\text{sed-water}}$  can be determined based on the equation below and by using the default values from EU 2018a:

$$K_{\text{sed-water}} = F_{\text{air}_{\text{sed}}} \times K_{\text{air-water}} + F_{\text{water}_{\text{sed}}} + F_{\text{solid}_{\text{sed}}} \times (K_{p_{\text{sed}}} / 1000) \times RHO_{\text{solid}} =$$
$$K_{\text{sed-water}} = 0 + 0.8 + 0.2 \times (10.8 / 1000) \times 2500 = 6.2 \text{ m}^3\text{m}^{-3}$$

The QS for sediment can be determined based on the following equations, and by applying AA-EQS<sub>freshwater</sub> of 0.003 µg/l and AA-EQS<sub>saltwater</sub> of 0.0006 µg/l:

$$QS_{\text{sediment, freshwater}} = (K_{\text{sed-water}} / RHO_{\text{sed}}) \times QS_{\text{water}} \times 1000 = (6.2 / 1300) \times 0.003 \times 1000 = 0.014 \text{ } \mu\text{g/kg}_{\text{wet weight}}$$

$$QS_{\text{sediment, saltwater}} = (K_{\text{sed-water}} / RHO_{\text{sed}}) \times QS_{\text{water}} \times 1000 = (6.2 / 1300) \times 0.0006 \times 1000 = 0.0027 \text{ } \mu\text{g/kg}_{\text{wet weight}}$$

The QS<sub>sediment</sub> in wet weight can be converted to dry weight by applying the conversion factor of 2.6 (EU 2018a):

$$QS_{\text{sediment, freshwater}} = 0.014 \text{ } \mu\text{g/kg}_{\text{wet weight}} \times 2.6 = 0.04 \text{ } \mu\text{g/kg}_{\text{dry weight}}$$

$$QS_{\text{sediment, saltwater}} = 0.0027 \text{ } \mu\text{g/kg}_{\text{wet weight}} \times 2.6 = 0.007 \text{ } \mu\text{g/kg}_{\text{dry weight}}$$

$$\begin{aligned} QS_{\text{sediment, freshwater}} &= \mathbf{0.04 \text{ } \mu\text{g/kg dry weight (5\% OC)}} \\ &= \mathbf{0.8 \text{ } \mu\text{g/kg dry weight} \times f_{oc}} \\ QS_{\text{sediment, saltwater}} &= \mathbf{0.007 \text{ } \mu\text{g/kg dry weight (5\% OC)}} \\ &= \mathbf{0.14 \text{ } \mu\text{g/kg dry weight} \times f_{oc}} \end{aligned}$$

### QS for secondary poisoning

Data on birds and mammal were taken primarily from FMC 2019 and ECOTOX. As malathion is most likely bio-diluted the QS<sub>biota-secondary poisoning</sub> was calculated for bivalves. All NOEC values for birds and mammal described in Bilag A are energy normalized by applying method B from EU 2018a, assuming that the animals food items contain an energy content of 15.1 kJ/g<sub>dry weight</sub> and a moisture fraction of 8% (table 8 in EU 2018a). Afterwards, all NOEC values were energy normalized to concentrations in mussels, assuming that mussels contain an energy content of 19 kJ/g<sub>dry weight</sub> and a moisture fraction of 92% (table 7 in EU 2018a) (se Bilag B).

The lowest NOEC presented in Bilag B is 2.7 mg/kg mussels (wet weight) and an AF of 10 is applied.

$$QS_{\text{sec. pois.}} = \mathbf{2.7 \text{ mg/kg mussels} / 10 = 0.27 \text{ mg/kg mussels wet weight}}$$

### QS for human health

The QS<sub>biota-human health</sub> was based on an ADI of 0.03 mg/kg bw per day and determined from the following equation:

$$QS_{\text{human health}} = 0.2 \times \text{ADI} / 0.00163 = 0.2 \times 30 / 0.00163 = 3681 \text{ } \mu\text{g/kg fishery products} \approx$$

$$QS_{\text{human health}} = \mathbf{3.7 \text{ mg/kg wet weight Fish, Fishery products}}$$

**Q<sub>Swater</sub> based on Q<sub>Sec. pois.</sub> and Q<sub>Shuman health</sub>**

The corresponding concentration in water was calculated based on the lowest value for Q<sub>Swater, sec.pois.</sub> = 0.27 mg/kg and a BCF of 100 l/kg:

$$Q_{\text{Swater, biota}} = 0.27 \text{ mg/kg} / 100 \text{ l/kg} = 0.0027 \text{ mg/l} \approx 2.7 \text{ } \mu\text{g/l}$$

The QS is above the AA-EQS for both fresh- and saltwater, and the derived AA-EQS is considered to protect against secondary poisoning and human health.

In conclusion, the following EQS for the aquatic environment have been derived for malathion:

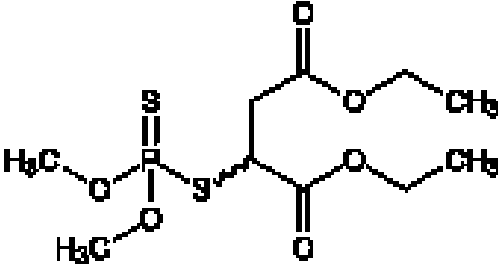
<b>AA-EQS<sub>freshwater</sub></b>	<b>= 0.003 <math>\mu\text{g/l}</math></b>
<b>AA-EQS<sub>saltwater</sub></b>	<b>= 0.0006 <math>\mu\text{g/l}</math></b>
<b>MAC-EQS<sub>freshwater</sub></b>	<b>= 0.4 <math>\mu\text{g/l}</math></b>
<b>MAC-EQS<sub>saltwater</sub></b>	<b>= 0.4 <math>\mu\text{g/l}</math></b>
<b>Q<sub>Ssediment, freshwater</sub></b>	<b>= 0.04 <math>\mu\text{g/kg}</math> dry weight (5% OC)</b> <b>= 0.8 <math>\mu\text{g/kg}</math> dry weight x f<sub>oc</sub></b>
<b>Q<sub>Ssediment, saltwater</sub></b>	<b>= 0.007 <math>\mu\text{g/kg}</math> dry weight (5% OC)</b> <b>= 0.14 <math>\mu\text{g/kg}</math> dry weight x f<sub>oc</sub></b>
<b>Q<sub>Sec. pois.</sub></b>	<b>= 0.27 mg/kg mussels wet weight</b>
<b>Q<sub>Shuman health</sub></b>	<b>= 3.7 mg/kg wet weight Fish, Fishery products</b>
<b>Q<sub>Swater, biota</sub></b>	<b>= 2.7 <math>\mu\text{g/l}</math></b>

# 1 Indledning

Identiteten af malathion fremgår af tabel 1.1.

Malathion er et acetylcholinesterasehæmmende insekti- og acaricid. Stoffet er godkendt til visse brug i EU.

Tabel 1.1. Identitet af malathion

IUPAC navn	diethyl 2-dimethoxyphosphinothioylsulfanylbutanedioate
Strukturformel	
CAS nr.	121-75-5
EINECS nr.	204-497-7
Kemisk formel	C <sub>10</sub> H <sub>19</sub> O <sub>6</sub> PS <sub>2</sub>
SMILES	CCOC(=O)CC(C(=O)OCC)SP(=S)(OC)OC
Harmoniseret klassificering	Acute Tox. 4; H302 (Farlig ved indtagelse) Skin Sens. 1; H317 (Kan forårsage allergisk hudreaktion) Aquatic Acute 1; H400 (Meget giftigt for vandlevende organismer) Aquatic Chronic 1; H410 (Meget giftigt med langvarige virkninger for vandlevende organismer)
Selvklassificering	



## 2 Fysisk kemiske egenskaber

De fysisk kemiske egenskaber for malathion fremgår af tabel 2.1.

Tabel 2.1. Fysisk kemiske egenskaber for malathion

Parameter	Værdi	Reference
Molekylvægt, $M_w$ ( $\text{g}\cdot\text{mol}^{-1}$ )	330,4	EFSA 2009
Smeltepunkt, $T_m$ ( $^{\circ}\text{C}$ )	< -20	EFSA 2009
Kogepunkt, $T_b$ ( $^{\circ}\text{C}$ )	Desintegrerer ved 174 grader	EFSA 2009
Damptryk, $P_v$ (Pa)	$4,5\cdot 10^{-4}$	EFSA 2009
Henry's konstant, H ( $\text{Pa m}^3 \text{mol}^{-1}$ ) <sup>a</sup>	$1\cdot 10^{-3}$	EFSA 2009
Vandopløselighed, $S_w$ ( $\text{g}\cdot\text{L}^{-1}$ )	0,148 <sup>a</sup> 0,143-0,145 <sup>b</sup>	EFSA 2009, FMC 2019: 1994; 076 FYF PubChem
Dissociationskonstant, $\text{pK}_a$	Dissocierer ej	EFSA 2009
Octanol/vand fordelingskoefficient, $\log K_{ow}$	2,75 2,36-2,86	EFSA 2009, FMC 2019 PubChem
Sediment/vand fordelingskoefficient, normaliseret til organisk karbon, $K_{oc}$ ( $\text{L}\cdot\text{kg}^{-1}$ ). $K_{oc}$ for jord er brugt	151-308	EFSA 2009

<sup>a</sup> Ved 25°C

<sup>b</sup> Ved 20°C

## 3 Skæbne i miljøet

### 3.1 Nedbrydelighed

Omdannelse af moderstoffet i jord (primær nedbrydelighed) har en halveringstid på  $DT_{50}$  mellem 0,08 dag og 1,87 dag (FMC (2019), Document M-CA Section 7).

Ifølge Dortland (1980) var halveringstiden i vand omkring 2 dage, hvilket er i overensstemmelse med forsøg indleveret til pesticidgodkendelsen (Dobson & Petts (2019) i FMC, 2019), der fandt en halveringstid på 2,24 dage.

De vigtigste omdannelsesprodukter af malathion i vand er malathion mono-carboxylsyre (MMCA) og malathion di-carboxylsyre (MDCA). Deres giftighed overfor *Lepomis macrochirus* og *Daphnia magna* er flere tusinde gange mindre end malathions, så de bidrager efter alt at dømme kun meget lidt til den samlede giftighed, selvom de er i vandet længere end malathion. (FMC 2019: Document MCA, section 7 og 8).

I studier med både vand og sediment ser halveringstiden for malathion i vandet ud til at være mindre end et døgn, mens MMCA har en halveringstid på 1,4 – 2,9 døgn og MDCA har en halveringstid på 5,5 døgn (Knoch 2001 og Cooke 2019).

### 3.2 Bioakkumulering

Som det fremgår af tabel 2.1, varierer de angivne  $\log K_{ow}$  mellem 2,36 og 2,86.

Upubliceret (1994, dokument 076 FYF, refereret i FMC, 2019) angiver at, BCF for hel fisk er 95, og at kinetisk<sup>1</sup> BCF er 103. Forsøget er udført med *Lepomis macrochirus*. Forsøget er også citeret, og altså vurderet og accepteret, i EFSA 2009.

Den danske QSAR database (Danish (Q)SAR Database (dtu.dk))(se datablad i bilag C) forudsiger en BCF mellem 16,8 – 25,4 og en BAF mellem 24,4 – 26,4, afhængig af den valgte model. De målte værdier har større vægt end QSAR, men resultaterne fra QSAR peger ikke på, at BCF og BAF skulle være større end de målte værdier.

I rotter omsættes malathion hurtigt. Stoffet udskilles således med over 90% på 24 timer.

Det er derfor usandsynligt, at malathion opkoncentreres i fødekæden, man vil snarere forvente det modsatte.

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<sup>1</sup> Kinetisk BCF = koncentrationen i fisk / udskillelsesrate i fisk.

Martinez-Tabche et al. 2002 angiver en BCF i en snegl (*Stagnicola sp.*) på 45 l/kg. Det angives dog, at det er koncentrationen i dyrene divideret med koncentrationen i sedimentet. Det er derfor usikkert, hvor sammenligneligt det er med BCF i fisk, og énheden l/kg er i øvrigt tvivlsom.

Man kan dog sige, at der ikke umiddelbart er noget, der tyder på en større BCF i snegle end i fisk, og efter 3 dage var malathion stort set ude af dyrene.

### 3.3 Naturlig forekomst

Der er ingen naturlig forekomst af malathion.

# 4 Toksicitetsdata

## 4.1 Toksicitet over for vandlevende organismer

De akutte og kroniske datasæt ses i bilag A. De værdier, der er brugt i SSD (Species Sensitivity Distribution) analyserne er i bilaget mærket med fed skrift.

Som udgangspunkt er data taget fra FMC (2019) og US EPAs database ECOTOX. Der er meget store datamængder i ECOTOX databasen, specielt vedrørende akut toksicitet.

Den første sortering af kroniske data fra ECOTOX er sket ud fra oplysninger givet om varighed af test, effekttyper (endpoints), om der er analytisk bekræftelse af testkoncentrationerne og om der er anvendt en formulering eller aktivstoffet. Generelt er studier af for kort varighed i forhold til standardtest udeladt, og effekttyper, der vanskeligt kan relateres til effekter på populationer, er heller ikke medtaget. Studier, hvor der anvendes en formulering, og koncentrationerne af stoffet ikke er analytisk målt, er også udeladt. Studier med kun en eller to testkoncentrationer, udover kontrollen, er ikke medtaget. Studier er dog blevet accepteret, hvor stoffet ikke er målt, men aktivstof har været anvendt, og testopløsningerne er blevet fornyede gennem forsøget.

Så vidt muligt er alle referencerne til de kroniske test, som ikke er med i FMC (2019), skaffet og kvalitetsvurderet efter CRED (Moermond et al, 2016). Kun studier med et troværdighedsindeks (RI) på 1 eller 2 er medtaget i beregningerne af vandkvalitetskriterierne.

Studier accepteret i FMC (2019) er accepteret uden yderligere CRED-vurdering.

Der er meget store mængder akutte data i ECOTOX (1795). Udvalgelsen af akutte data er sket efter de samme principper som for de kroniske data. Dog er manglende målinger accepteret i op til 2 dage, da halveringstiden skønnes at være højst 2 dage og da der er tale om akutte virkninger. For at få så mange af arterne repræsenteret i datasættet er mange 2-dages test blevet accepteret, hvor standardtestens varighed f.eks. er 4 dage. Til gengæld er den laveste af værdierne for hver art blevet valgt. Efter første grovsortering var der 311 værdier. For de laveste og de højeste værdier i ECOTOX er referencerne, så vidt muligt, fremskaffet og CRED vurderede, 47 i alt, dækkende 58 af effektkoncentrationerne. De øvrige effektkoncentrationer er ikke troværdighedsvurderede.

Hvor der er flere værdier for den samme art fra andre kilder end ECOTOX er alle værdierne medtaget, forudsat de ser ud til at være foretaget under de samme betingelser og vedrører samme typer effekter. I det endelige datasæt er det geometriske gennemsnit så anvendt.

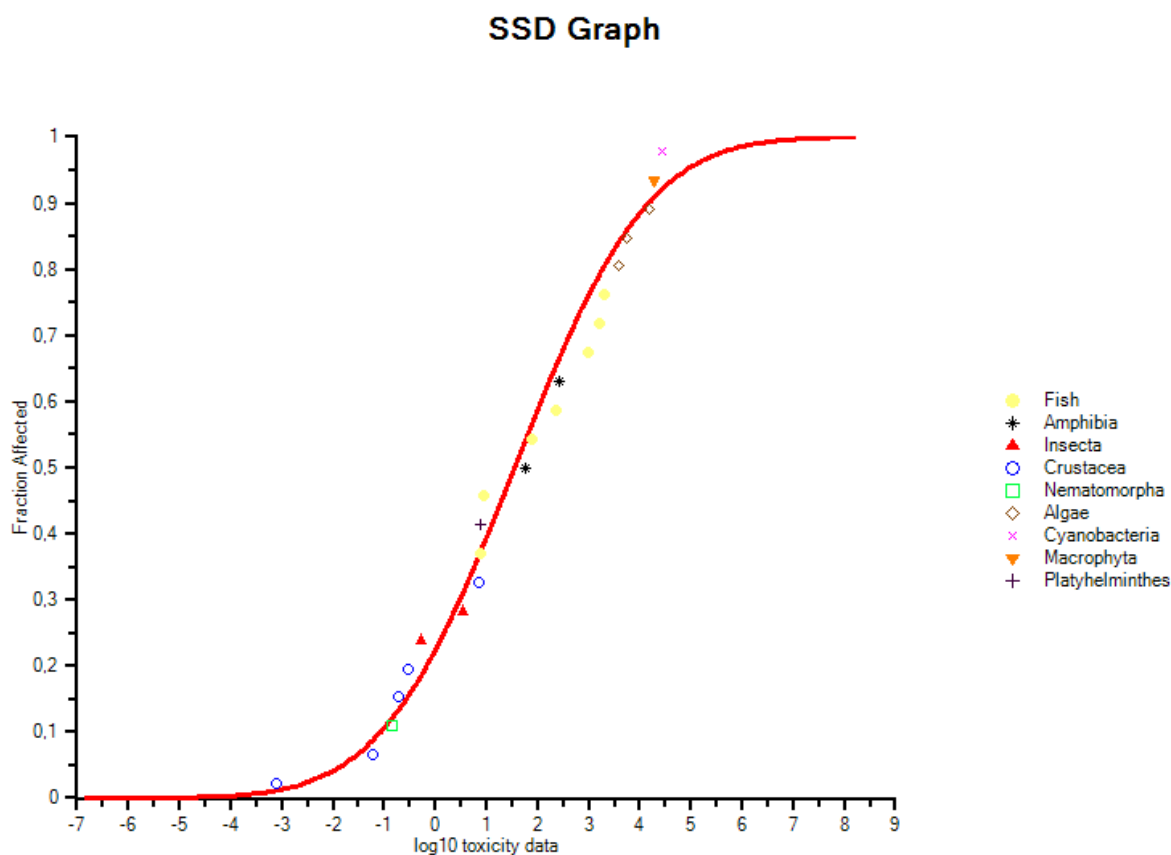
Der er, statistisk set, ikke signifikant forskel mellem den akutte giftighed overfor ferskvandsorganismer og saltvandsorganismer (t-test,  $P = 0,076$ ;  $N_1 = 55$ ,  $N_2 = 271$ ). Det skal her bemærkes, at det er et meget stort datamateriale, og at selv ganske små forskelle vil resultere i statistisk signifikans.

Der er heller ikke statistisk signifikant forskel mellem kronisk giftighed overfor fersk- og saltvandsorganismer (t-test,  $P = 0,613$ ;  $N_1 = 4$ ,  $N_2 = 21$ ).

For både akut og kronisk toksicitet gælder det derfor at datasættene for fersk- og saltvand kan lægges sammen.

Kronisk toksicitet:

Det kroniske datasæt indeholder værdier for 23 arter repræsenterende 9 overordnede systematiske grupper, og en artsfølsomhedsanalyse (SSD) kan derfor udføres. Data er analyseret med programmet ETx2, der kan hentes på RIVMs hjemmeside ([ETx | Risico's van stoffen \(rivm.nl\)](#)). Værdierne, der indgår i SSD-analysen er markeret med fed i bilag A.



$HC_5$ -kronisk, fersk+salt = 0,0141  $\mu\text{g/l}$ , 90% CI = 0,00063  $\mu\text{g/l}$  – 0,126  $\mu\text{g/l}$ . Standardafvigelsen af de log<sub>10</sub> transformerede data = 2,04. Alle tre ”goodness of fit” test i ETx accepterer hypotesen om log-normalfordeling på alle de listede signifikansniveauer.

Akut toksicitet:

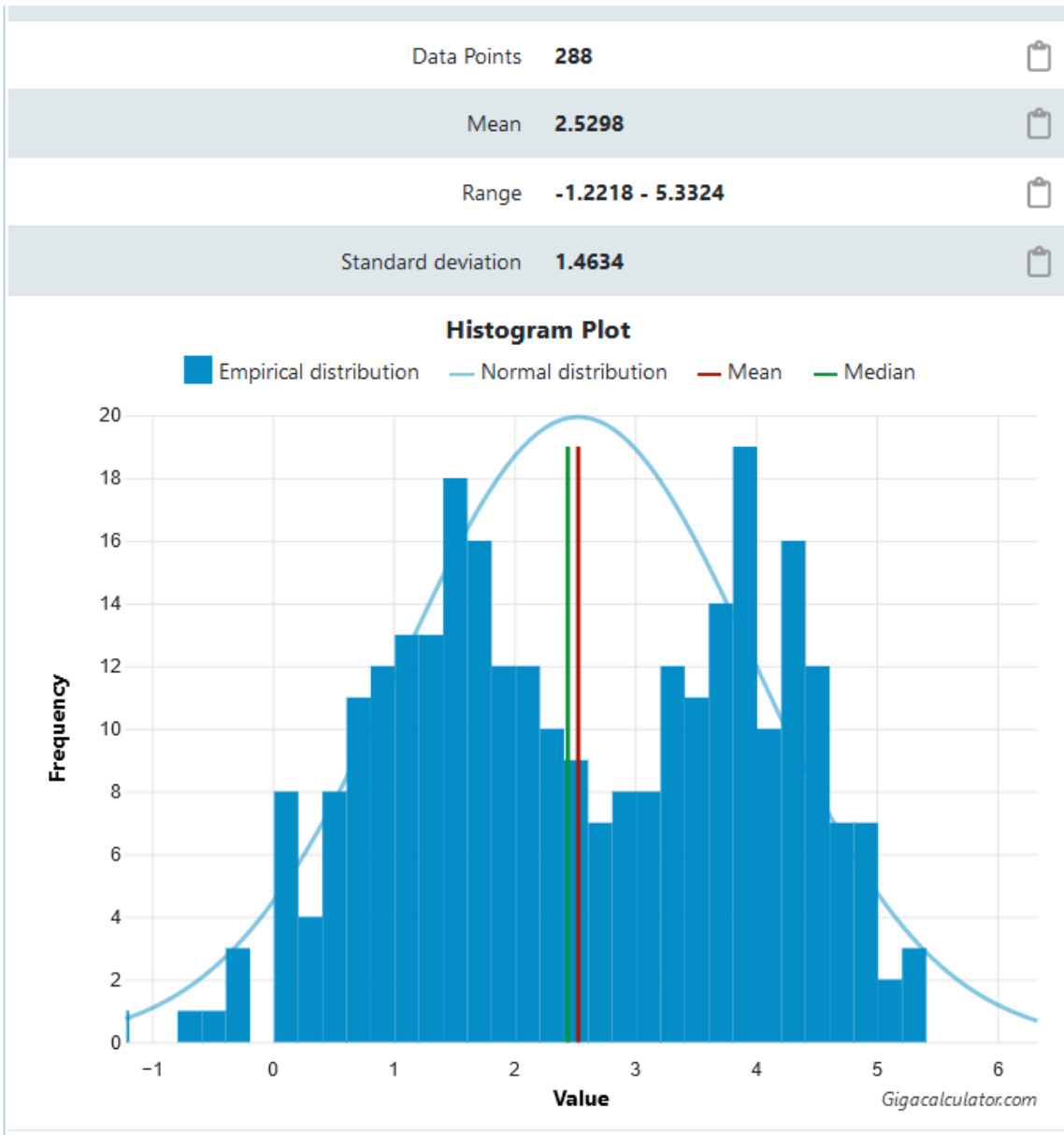
Det akutte datasæt indeholder  $EC_{50}$  værdier for 272 arter repræsenterende 16 systematiske hovedgrupper og en SSD kan derfor bruges. Værdierne, der indgår i SSD-analysen er markeret med fed i bilag A.

$HC_5$  for det samlede materiale er  $0,88 \mu\text{g/l}$  med en standardafvigelse (SD) for de  $\log_{10}$  transformerede værdier på 1,47.

Data fordeler sig meget klart i to grupper (se figur nedenfor) og data er klart ikke normalfordelte. Der udføres derfor en SSD analyse med data repræsenterende den nedre halvdel af fordelingen.

$HC_5$  for den nedre halvdel af fordelingen er  $1,2 \mu\text{g/l}$  med nedre og øvre 90% konfidensgrænser på  $0,8 \mu\text{g/l} - 1,8 \mu\text{g/l}$  samt en SD på 0,80 (N = 159). Alle tre "goodness of fit" programmer i ETx accepterer lognormalfordeling på alle signifikansniveauerne.

Dette datasæt omfatter 159 arter repræsenterende syv overordnede systematiske grupper (hjuldyr, muslinger, snegle, krebsdyr, insekter, fisk og padder). De grupper, som må forventes at indeholde de mest følsomme arter, krebsdyr og insekter er rigt repræsenterede. Der er således 53 arter krebsdyr og 70 arter insekter. Krebsdyrene og insekterne kan underopdeles i meget forskellige grupper med hensyn til generel økologi og f.eks. fødesøgningsstrategier. Krebsdyrene omfatter: Branchiopoda (bladfødder), Amphipoda, Decapoda, Ostracoda (muslingekrebs) og Isopoda. Insekterne omfatter: Tæger, døgnfluer, guldsmede, slørvinger, vårfluer tovinger og biller.



#### 4.2 Toksicitet over for sedimentlevende organismer

Der foreligger  $EC_{50}$ -værdier fra test i vand uden sediment med en række organismer, som normalt lever i eller på sediment. Disse er bl.a.:

Visse Chironomus arter med  $EC_{50} < 1 \mu\text{g/l}$

Visse Gammarus arter med  $EC_{50} < 1$  “

Hyalella sp. Med  $EC_{50} = 0,06$  “

Der er således en klar indikation på stor giftighed overfor sedimentlevende organismer.

Der er ikke fundet brugbare data for sediment ved søgning i FMC 2019, ECOTOX databasen, Agris, Biological Abstracts, GeoScience World, PubMed og Google Scholar.

Der er dog én reference, som giver en LC<sub>50</sub> værdi (Martínez-Tabche 2001) for *Limnodrilus hoffmeisteri* (en oligochaet) på 0,269 mg/kg sediment (2,25% OC). En LC<sub>50</sub> er ikke relevant og studiets opfyldelse af validitetskriterierne kan ikke vurderes på grund af manglende oplysninger.

#### 4.3 Toksicitet over for pattedyr og fugle

Giftigheden overfor fugle og pattedyr er angivet i bilag A.

Data er hentet fra FMC (2019) og ECOTOX databasen.

Malathion betragtes ikke som biomagnificerende, og det kritiske fødeemne for vilde fugle og pattedyr er derfor invertebrater som f.eks. muslinger.

Da stoffet ikke er specielt hydrofobt (vandopløselighed er 143 mg/l), og da lipidkoncentrationen i muslinger er lav (omkr. 1%, EU 2018b) beregnes koncentrationen i muslinger som mg/kg tørvægt.

NOEC i muslinger for de enkelte arter er givet i bilag B. Hvor der er flere NOEC-værdier for en art er det geometriske gennemsnit brugt.

Laveste NOEC i muslinger er 2,7 mg/kg<sub>vådvægt</sub> musling (bløddele), som anvendes i udledningen af biotakvalitetskriteriet for sekundær forgiftning.

#### 4.4 Toksicitet over for mennesker

For menneskers sundhed er malathion klassificeret med H302 (giftigt, hvis slugt) og H317 (hudsensibiliserende) (Annex VI til CLP, ATP17:

[https://echa.europa.eu/documents/10162/17218/annex\\_vi\\_clp\\_table\\_atp17\\_en.xlsx/4dcec79c-f277-ed68-5e1b-d435900dbe34?t=1638888918944&download=true](https://echa.europa.eu/documents/10162/17218/annex_vi_clp_table_atp17_en.xlsx/4dcec79c-f277-ed68-5e1b-d435900dbe34?t=1638888918944&download=true)).

ADI (Acceptable Daily Intake) = 0,03 mg/kg lgv/d (EFSA 2009). Værdien er baseret på langtids studier med rotter.

Jævnfør EU (2018b) er ADI 0,03 mg/kg lgv/d svarende til 30 µg/kg lgv d



## 5 Andre effekter

Der er ikke fundet oplysninger om andre typer af effekter.

## 6 Udledning af vandkvalitetskriterium

Kvalitetskriterierne er fastsat i overensstemmelse med EU's Guidance Document no. 27: Technical Guidance for Deriving Environmental Quality Standards (EU, 2018a).

For beregning af VKK er retningslinjerne for fastsættelse af usikkerhedsfaktorer (UF), der skal anvendes på HC<sub>5</sub>-værdier fra SSD analyser begrænset til, at faktoren skal være mellem 5-1, samt at der er en række aspekter, som man skal tages højde for (f.eks. antal arter, varians m.m., EU 2018a, side 45 afsnit 3.3.1.2). Udgangspunktet er en UF = 5, når betingelserne for at udføre en SSD analyse lige akkurat er opfyldt. Med flere og bedre data kan UF reduceres. Ved beregning af KVKK er udgangspunktet en UF = 10.

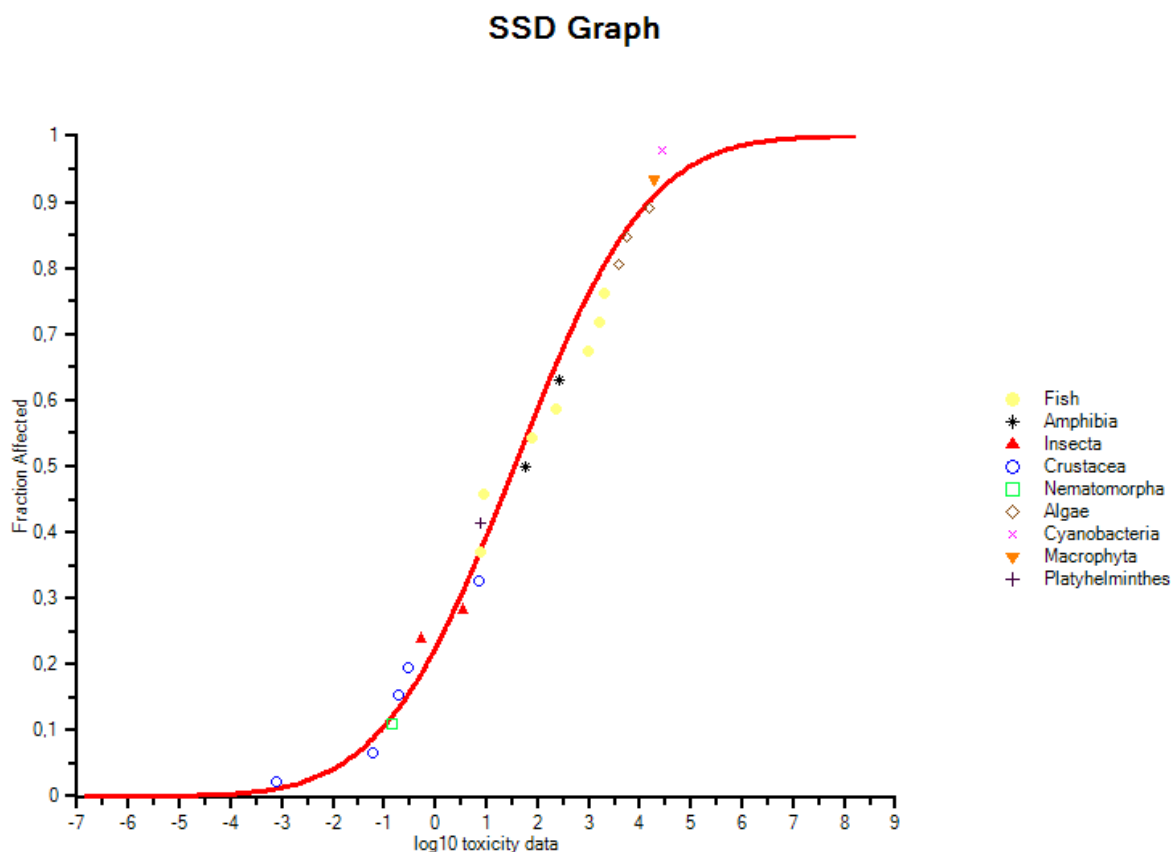
UF værdierne 5-1 blev fastlagt i 2001 førend man var helt på det rene med hvilken grad af effekt NOEC svarede til og hvilken effektstørrelse EC<sub>x</sub> skulle repræsentere. Da det i dag er klart, at NOEC i gennemsnit svarer til omkr. 10% effekt og, at man foretrækker at bruge EC<sub>10</sub> giver det ikke længere mening at anvende en UF = 1.

I en årrække har Miljøstyrelsen anvendt følgende ”tommelfingerregel” i nedenstående tabel ved valg af UF på HC<sub>5</sub> (VKK):

<b>Antal arter</b>		<b>Antal overordnede systematiske grupper</b>	<b>Usikkerhedsfaktor</b>
<b>10-14</b>	og	<b>8-9</b>	<b>5</b>
<b>15-20</b>	eller	<b>10-11</b>	<b>4</b>
<b>21-30</b>	eller	<b>12-14</b>	<b>3</b>
<b>≥31</b>	eller	<b>≥15</b>	<b>2</b>

### 6.1 Vandkvalitetskriterium (VKK)

HC<sub>5</sub> = 0,014 µg/l og standardafvigelsen af de log<sub>10</sub> transformerede værdier er 2,04. Anderson-Darling, Kolmogorov-Smirnov og Kramer von Mises tests for normalfordeling accepterer lognormalfordelingen på alle signifikansniveauerne.



Der er 23 arter og 9 overordnede systematiske grupper, og som udgangspunkt vælges en usikkerhedsfaktor (UF) på 3.

Variansen er dog stor ( $SD = 2,04$ ) og i det akutte datasæt er der tretten arter, der er mere følsomme end *Ceriodaphnia*, som er den mest følsomme kroniske art. Tolv af disse tretten arter er ikke repræsenteret i det kroniske datasæt. Af de nævnte tolv arter er 10 krebsdyr eller insekter, grupper som må forventes at være blandt de mest følsomme.

Derfor ville man nok vælge en UF på 4.

$VKK_{\text{ferskvand}}$  ville derfor blive  $0,014 \mu\text{g/l} / 4 = 0,0035 \mu\text{g/l}$

Det bemærkes dog, at denne værdi er større end  $EC_{10}$  for den mest følsomme af de testede arter,  $0,0008 \mu\text{g/l}$  for *Ceriodaphnia dubia*, samt at det akutte datasæt indeholder tretten arter, der er mere følsomme end *C. dubia*, og tolv af disse arter er ikke repræsenteret i det kroniske datasæt.

Det vælges derfor ikke at sænke UF men beholde UF på 5. Dette resulterer i en værdi på:

$VKK_{\text{ferskvand}} = 0,014 \mu\text{g/l} / 5 = 0,0028 \mu\text{g/l} \approx 0,003 \mu\text{g/l}$

Denne værdi er stadig 3,75 gange større end EC<sub>10</sub> for *Ceriodaphnia dubia*, men tyve gange mindre end værdien for *Daphnia magna*, som er den næstmest følsomme i det kroniske datasæt.

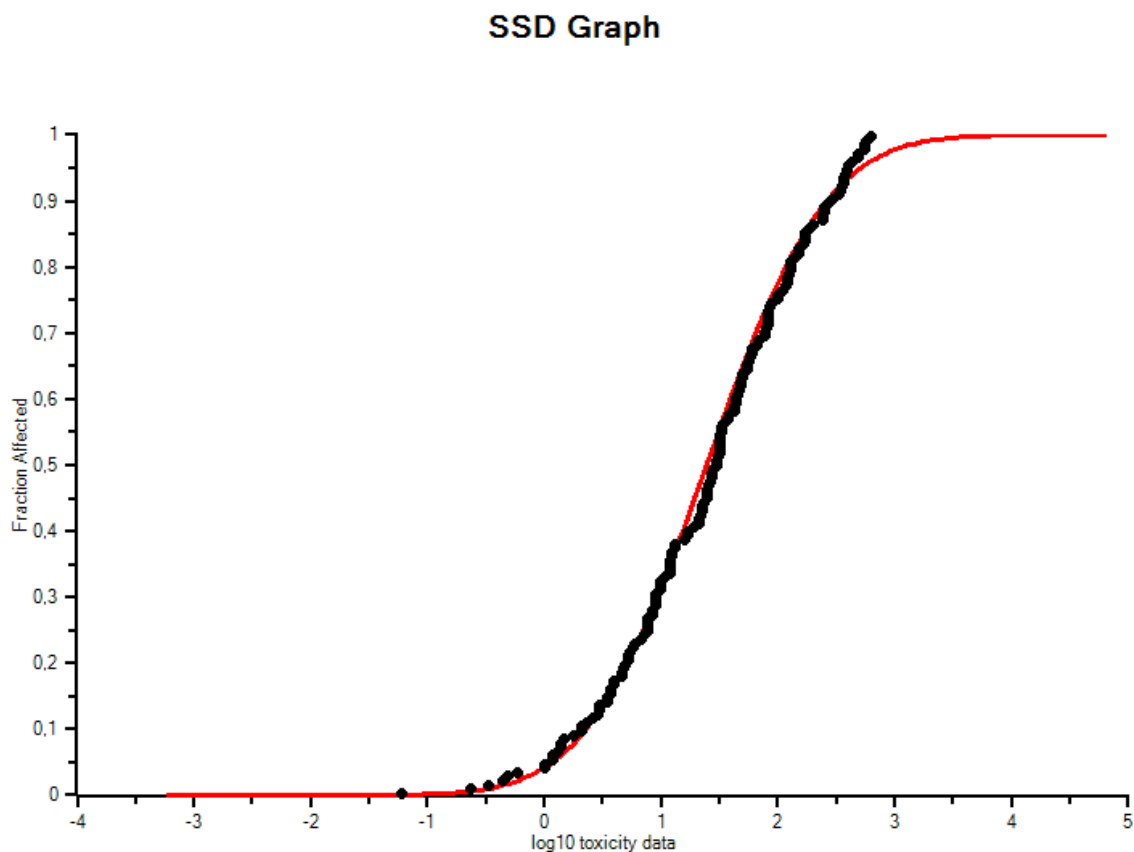
I det kroniske datasæt for saltvand er der kun én speciel repræsentant for det marine miljø, krabben *Cancer magister*. Der skal derfor anvendes en ekstra usikkerhedsfaktor på 5 ved beregning af VKK for det marine miljø.

$$\text{VKK}_{\text{saltvand}} = 0,0028 \mu\text{g/l} / 5 = \mathbf{0,00056 \mu\text{g/l}} \approx \mathbf{0,0006 \mu\text{g/l}}$$

## 6.2 Korttidsvandkvalitetskriterium (KVKK)

Da fordelingen af EC<sub>50</sub> værdier er to-toppet anvendes HC<sub>5</sub> for den nedre gruppe i fordelingen.

HC<sub>5-akut</sub> = 1,2 for den nedre del af fordelingen, og SD af de log<sub>10</sub> transformerede tal = 0,80. En lognormal fordeling er accepteret i alle tilfælde.



I det akutte marine datasæt er der adskillige arter, der, jævnfør EU vejledningen (EU 2018a), kan regnes for specielle repræsentanter for saltvand inden for de taksonomiske grupper hjuldyr, muslinger og pighuder. Der skal derfor ikke føjes en ekstra usikkerhedsfaktor til beregning af KVKK for saltvand. KVKK for saltvand bliver lig KVKK for ferskvand.

LC<sub>50</sub> datasættet er meget omfattende (272 arter, 15 overordnede grupper) og omfatter mange arter, som må forventes at være blandt de mest følsomme, nemlig insekter og krebsdyr. Der er således 53 arter krebsdyr og 70 arter insekter, og materialet omfatter 6 klasser af krebsdyr og 7 insektordener. Ovennævnte HC<sub>5</sub> er for den ”nederste” gruppering af EC<sub>50</sub> værdierne. Denne gruppering omfatter 159 arter repræsenterende 7 overordnede systematiske grupper. Krebsdyrene og insekterne kan underopdeles i økologisk meget forskellige grupper, og betingelserne for at udføre en SSD analyse er klart til stede.

Som udgangspunkt skal man, ifølge EU vejledningen (EU 2018a), anvende en UF på 10. Men, med det meget omfattende datamateriale og omfattende repræsentation af særligt følsomme grupper, foreslås det at anvende en UF på 3, også selvom variansen er stor.

$$\text{KVKK}_{\text{ferskvand}} = \text{KVKK}_{\text{saltvand}} = 1,2 \mu\text{g/l} / 3 = \mathbf{0,4 \mu\text{g/l}}$$

Der er 4 arter, som har en lavere EC<sub>50</sub> end denne værdi: *Hyalella azteca* (0,06 μg/l), *Jordanella floridae* (0,24 μg/l), *Gammarus pulex* (0,33 μg/l) og *G. fasciatus* (0,48 μg/l). Dette kunne være et argument for, at anvende en større UF, men dels er datamængden særdeles omfattende, dels udgør disse arter kun 1,5% af alle de testede arter og 2,5% af den ”nedre gruppe”. Endvidere lægges der større vægt på kroniske effekter, som relaterer til vedvarende udledninger.

### 6.3 Kvalitetskriterium for sediment (SKK)

Den store akutte giftighed (test i vand) over for en række organismer, der normalt lever i/på sediment er en klar indikation på, at et SKK kan være relevant, og et kvalitetskriterie for sediment bør udarbejdes.

EqP-metoden (EU 2018a) anvendes, da der ikke er troværdige tilgængelige kronisk effektdata fra sedimentstudier.

Der anvendes et EU standard sediment med et organisk kulstof (OC)-indhold på 0,05 (5%) og en K<sub>oc</sub>-værdi. Det geometriske gennemsnit af K<sub>oc</sub> værdierne angivet i tabel 2.1 er 216 l/kg.

Fordelingskoefficienten mellem fast stof og vand i sediment, K<sub>p<sub>sed</sub></sub> bestemmes ved følgende formel:

$$K_{p_{\text{sed}}} = F_{OC_{\text{sed}}} \times K_{OC} = 0,05 \times 216 \text{ l/kg} = 10,8 \text{ l/kg}$$

Fordelingskoefficienten mellem sediment og vand, K<sub>sed-vand</sub> kan hernæst bestemmes ved følgende formel, med de nedenfor angivne ”default” værdier fra EU vejledningen (EU 2018a):

$$K_{\text{sed-vand}} = F_{\text{luft}_{\text{sed}}} \times K_{\text{luft-vand}} + F_{\text{vand}_{\text{sed}}} + F_{\text{fast}_{\text{sed}}} \times (K_{p_{\text{sed}}} / 1000) \times RHO_{\text{fast}}$$

”Default” værdier fra EU vejledningen:

F <sub>luft<sub>sed</sub></sub> x K <sub>luft-vand</sub> :	0
F <sub>vand<sub>sed</sub></sub> :	0,8 m <sup>3</sup> m <sup>-3</sup>
F <sub>fast<sub>sed</sub></sub> :	0,2
RHO <sub>fast</sub> :	2500 kg*m <sup>-3</sup>
RHO <sub>sed</sub> :	1300 kg*m <sup>-3</sup>

$$K_{\text{sed-vand}} = 6,2 \text{ m}^3 \text{m}^{-3}$$

SKK for ferskvand og saltvand kan bestemmes ved anvendelse af nedenstående formler, hvor VKK på 0,003 µg/l og 0,0006 µg/l for hhv. ferskvand og saltvand anvendes:

$$\text{SKK}_{\text{vådvægt, ferskvand}} = (K_{\text{sed-vand}} / \text{RHOSed}) \times \text{VKK}_{\text{ferskvand}} \times 1000 = (6,2 / 1300) \times 0,003 \times 1000 = 0,014 \text{ µg/kg}_{\text{vådvægt}}$$

Faktor til konvertering til tørvægt:  $\text{RHOSed} / \text{Ffast}_{\text{sed}} \times \text{RHO}_{\text{fast}} = 2,6$

$$\text{SKK}_{\text{tørvægt, ferskvand}} = \text{SKK}_{\text{vådvægt}} \times 2,6 = 0,014 \text{ µg/kg}_{\text{vådvægt}} \times 2,6 = \mathbf{0,04 \text{ µg/kg}_{\text{tørvægt}} \text{ (5\% OC)}}$$

$$\text{SKK}_{\text{tørvægt, ferskvand}} = 0,04 / 0,05 \text{ µg/kg} = \mathbf{0,80 \text{ µg/kg}_{\text{tørvægt}} \text{ x foc}}$$

$$\text{SKK}_{\text{vådvægt, saltvand}} = (K_{\text{sed-vand}} / \text{RHOSed}) \times \text{VKK}_{\text{saltvand}} \times 1000 = (6,2 / 1300) \times 0,0006 \times 1000 = 0,0027 \text{ µg/kg}_{\text{vådvægt}}$$

$$\text{SKK}_{\text{tørvægt, saltvand}} = 0,0027 \times 2,6 = \mathbf{0,0069 \text{ µg/kg}_{\text{tørvægt}} \approx 0,007 \text{ µg/kg}_{\text{tørvægt}} \text{ (5\% OC)}}$$

$$\text{SKK}_{\text{tørvægt, saltvand}} = 0,0069 / 0,05 = \mathbf{0,14 \text{ µg/kg}_{\text{tørvægt}} \text{ x foc}}$$

#### 6.4 Kvalitetskriterium for biota, sekundær forgiftning ( $\text{BKK}_{\text{sek.forgiftn.}}$ )

Da BCF er målt til omkring 100 bør der udarbejdes BKK for sekundær forgiftning (fødekæde effekt) (EU 2018a).

I rotter elimineres malathion med mere end 90% på 24 timer og alt i alt tyder det på TMF (trophic magnification factor) og BMF (bio-magnification factor) værdier er mindre end 1, og  $\text{BKK}_{\text{sek.forgiftn.}}$  bør sættes for muslinger snarere end fisk.

NOEC-værdierne angivet i bilag A er alle energinormaliserede (mg/kJ), idet det antages at dyrenes foder har et energiindhold på 15,1 kJ/g<sub>tørvægt</sub> og et vandindhold på 8% (tabel 8 i EU-vejledningen, EU 2018a). Det er EU vejledningens metode B i afsnit 4.4.5, der er benyttet ved beregningerne da effektværdierne er koncentrationer i føden.

NOEC-værdierne er derefter omregnet til koncentrationer i muslinger, idet det antages at muslinger har et energiindhold på 19 kJ/g<sub>tørvægt</sub> og et vandindhold på 92% jævnfør afsnit 4.4.6 i EU vejledningen (EU 2018a). Værdierne er præsenteret i bilag B.

I bilag A er alle NOAEL (mg/kg lgv/ d) værdier blevet omregnet til NOEC-værdier med en omregningsfaktor givet i tabel 21 i EU vejledningen, og LC<sub>50</sub>-værdien for fasan er omregnet til NOEC ved at dividere med 100 (tabel 9 i EU 2018a).

Laveste NOEC er 2,7 mg/kg musling (vådvægt, bløddele) (se bilag B) og der anvendes en usikkerhedsfaktor på 10.

$$\mathbf{BKK} = 2,7 \text{ mg/kg musling} / 10 = \mathbf{0,27 \text{ mg/kg musling}}$$

## 6.5 Kvalitetskriterium for human konsum af vandlevende organismer (HKK)

BCF er omkring 100, og sammenholdt med klassificeringen Acute Tox. 4 bør der udarbejdes kvalitetskriterium for menneskers sundhed, human konsum (HKK).

Der er ikke fundet danske eller EU grænseværdier for malathion i fiskeriprodukter, og grænseværdien beregnes derfor med følgende formel (EU 2018a), ved anvendelse af ADI på 30 µg/kg lgv/ d (EFSA 20009):

$$\text{HKK} = 0,2 \times \text{ADI} / 0,00163 = 0,2 \times 30 / 0,00163 = 3681 \text{ µg/kg fiskeriprodukt} \approx$$

$$\text{HKK} = 3,7 \text{ mg/kg fiskeriprodukt}$$

## 6.6 Vandkvalitetskriterium baseret på BKK<sub>sek.forgiftn.</sub> og HKK

Jævnfør EU vejledningen (EU, 2018a) skal der laves en tilbageregning fra biotakvalitetskriterierne (BKK<sub>sek.forgiftn.</sub> og HKK) til en vandkoncentration, for at se om vandkvalitetskriteriet fastsat for direkte effekter, også beskytter for sekundær forgiftning gennem fødekæden, samt beskytter mod forgiftning ved human konsum.

Der haves ingen BAF værdier, men for stoffer, der ikke opkoncentreres i fødekæden, svarer BAF stort set til BCF.

$$\text{BCF} \approx 100 \text{ l/kg}$$

$$\text{VKK}_{\text{vand, biota}} = \text{BKK} / \text{BCF}$$

$$\text{Laveste BKK eller HKK er BKK} = 0,27 \text{ mg/kg}$$

$$\text{VKK}_{\text{vand, biota}} = 0,27 \text{ mg/kg} / 100 \text{ l/kg} = 0,0027 \text{ mg/l} = 2,7 \text{ µg/l}$$

Denne værdi er højere end VKK for vand, og bliver derfor ikke udslagsgivende.

# 7 Konklusion

Følgende kvalitetskriterier for vandmiljøet er udregnet for malathion:

## Vandkvalitetskriterium

VKK<sub>ferskvand</sub> 0,003 µg/l

VKK<sub>saltvand</sub> 0,0006 µg/l

## Korttidsvandkvalitetskriterium

KVKK<sub>ferskvand</sub> 0,4 µg/l

KVKK<sub>saltvand</sub> 0,4 µg/l

## Sedimentkvalitetskriterium

SKK<sub>ferskvand</sub> 0,04 µg/kg<sub>tørvægt</sub> sediment (5% OC)  
0,8 µg/kg<sub>tørvægt</sub> x f<sub>oc</sub>

SKK<sub>saltvand</sub> 0,007 µg/kg<sub>tørvægt</sub> sediment (5% OC)  
0,14 µg/kg<sub>tørvægt</sub> x f<sub>oc</sub>

## Biotakvalitetskriterium, sekundær forgiftning

BKK<sub>sek.forgiftn.</sub> 0,27 mg/kg musling vådvægt

## Biotakvalitetskriterium, human konsum

HKK 3,7 mg/kg fiskeriprodukter,  
vådvægt



## 8 Referencer

Abbasi,S.A., and R. Soni 1991: Evaluation of Water Quality Criteria for Four Common Pesticides on the Basis of Computer-Aided Studies. <i>Indian J. Environ. Health</i> 33(1): 22-24
Achiorno,C.L., C. De Villalobos, and L. Ferrari 2009: Sensitivity of Preparasitic Stages of <i>Chordodes nobilii</i> (Gordiida, Nematomorpha) to Malathion. <i>Ecotoxicology</i> 18(5): 594-599
Aguilera,L., M.D.C. Marquetti, A. Navarro, and J. Bisset 1995: Effects of Three Organophosphorus Insecticides in the Reproductive Potential of <i>Culex quinquefasciatus</i> . <i>Mem. Inst. Oswaldo Cruz</i> 90(3): 411-413
Ahamad,I.K., K.V.R. Rao, and K.S. Swami 1978: Effect of Malathion on Enzyme Activity in Foot, Mantle and Hepatopancreas of Snail <i>Pila globosa</i> . <i>Indian J. Exp. Biol.</i> 16:258-260
Akarte,S.R., D.V. Muley, and U.H. Mane 1986: Effect of Cythion-Malathion 50 EC (W/W) on the Estuarine Clam, <i>Katylisia opima</i> (Gmelin). In: M.F.Thompson, R.Sarojini, and R.Nagabhushanam (Eds.), <i>Biology of Benthic Marine Organisms: Techniques and Methods as Applied to the Indian Ocean</i> , A.A.Balkema, Rotterdam, Netherlands:501-510
Aker,W.G., X. Hu, P. Wang, and H.M. Hwang 2008: Comparing the Relative Toxicity of Malathion and Malaoxon in Blue CatFisk, <i>Fish Ictalurus furcatus</i> . <i>Environ. Toxicol.</i> 23(4): 548-554
Aldenberg, T.; J.S. Jaworska et T.P. Traas 2002: Normal species sensitivity distributions and probabilistic ecological risk assessment. In <i>Species Sensitivity Distributions in Ecotoxicology</i> : 49-102. CRC Press, London
Ali,A. 1981: Laboratory Evaluation of Organophosphate and New Synthetic Pyrethroid Insecticides Against Pestiferous Chironomid Midges of Central Florida. <i>Mosq. News</i> 41(1): 157-161
Ali,A., and M.S. Mulla 1977: The IGR Diflubenzuron and Organophosphorus Insecticides Against Nuisance Midges in Man-Made Residential-Recreational Lakes. <i>J. Econ. Entomol.</i> 70(5): 571-577
Ali,A., and M.S. Mulla 1980: Activity of Organophosphate and Synthetic Pyrethroid Insecticides Against Pestiferous Midges in Some Southern California Flood Control Channels. <i>Mosq. News</i> 40:593-597
Ankley,G.T., J.R. Dierkes, D.A. Jensen, and G.S. Peterson 1991: Piperonyl Butoxide as a Tool in Aquatic Toxicological Research with Organophosphate Insecticides. <i>Ecotoxicol. Environ. Saf.</i> 21(3): 266-274
Ardo, J. 1974: K Otazke Akutnej Toxicity Niektorych Pesticidov. (Acute Toxicity of Some Pesticides). <i>Vodni Hospod.</i> 24(8): 222-224
Areekul,S. 1986: Toxicity to Fisk, Fishes of Insecticides Used in Paddy Fields and Water Resources. I. Laboratory Experiment. <i>Kasetsart J. Nat. Sci.</i> 20(2): 164-178
Arora,H.C., S.K. Shrivastava, and A.K. Seth 1971: Bioassay Studies of Some Commercial Organic Insecticides. Part II. Trials of Malathion with Exotic and Indigenous Carps. <i>Indian J. Environ. Health</i> 13(4): 300-306

Ashauer,R., A. Hintermeister, E. Potthoff, and B.I. Escher 2011: Acute Toxicity of Organic Chemicals to <i>Gammarus pulex</i> Correlates with Sensitivity of <i>Daphnia magna</i> Across Most Modes of Action. <i>Aquat. Toxicol.</i> 103(1/2): 38-45
Bahner,L.H., and D.R. Nimmo 1975: Methods to Assess Effects of Combinations of Toxicants, Salinity and Temperature on Estuarine Animals. <i>Trace Subst. Environ. Health</i> 9:169-177
Bailey,H.C., and D.H.W. Liu 1980: <i>Lumbriculus variegatus</i> , a Benthic Oligochaete, as a Bioassay Organism. <i>ASTM Spec. Tech. Publ.</i> :205-215
Bajet,C.M., A. Kumar, M.N. Calingacion, and T.C. Narvacan 2012: Toxicological Assessment of Pesticides Used in the Pagsanjan-Lumban Catchment to Selected Non-Target Aquatic Organisms in Laguna Lake, Philippines. <i>Agric. Water Manag.</i> 106:42-49
Bakry,F.A., W.S. Hasheesh, and S.A.H. Hamdi 2011: Biological, Biochemical, and Molecular Parameters of <i>Helisoma duryi</i> Snails Exposed to the Pesticides Malathion and Deltamethrin. <i>Pestic. Biochem. Physiol.</i> 101(2): 86-92
Bender,M.E., and J.R. Westman 1976: The Toxicity of Malathion and Its Hydrolysis Products to the Eastern Mudminnow, <i>Umbra pygmaea</i> (DeKay). <i>Chesapeake Sci.</i> 17(2): 125-128, 1976.
Beyers,D.W., T.J. Keefe, and C.A. Carlson 1994: Toxicity of Carbaryl and Malathion to Two Federally Endangered Fisk, Fishes, as Estimated by Regression and ANOVA. <i>Environ. Toxicol. Chem.</i> 13(1): 101-107
Birge,W.J., J.A. Black, and D.M. Bruser 1979: Toxicity of Organic Chemicals to Embryo-Larval Stages of Fisk, Fish. EPA-560/11-79-007, U.S.EPA, Washington, D.C.:60 p., 1979
Bogaerts,P., J. Bohatier, and F. Bonnemoy 2001: Use of the Ciliated Protozoan <i>Tetrahymena pyriformis</i> for the Assessment of Toxicity and Quantitative Structure-Activity Relationships of Xenobiotics: Comparison with the Microtox Test. <i>Ecotoxicol. Environ. Saf.</i> 49(3): 293-301
Boike,A.H.,Jr., C.B.,Jr. Rathburn, T.G. Floore, H.M. Rodriguez, and J.S. Coughlin 1989: Insecticide Tolerance of <i>Culex nigripalpus</i> in Florida. <i>Am. Mosq. Control Assoc. J.</i> 5(4): 522-528
Bonfanti,P., A. Colombo, F. Orsi, I. Nizzetto, M. Andrioletti, R. Bacchetta, P. Mantecca, U. Fascio, G. Vailati, and C. 2004: Comparative Teratogenicity of Chlorpyrifos and Malathion on <i>Xenopus laevis</i> Development. <i>Aquat. Toxicol.</i> 70(3): 189-200
Boumaiza,M., M.H. Ktari, and P. Vitiello 1979: Toxicity of Several Pesticides Used in Tunisia, for <i>Aphanius fasciatus</i> Nardo, 1827 (Pisces, Cyprinodontidae). <i>Arch. Inst. Pasteur Tunis</i> 56(3): 307-342
Brandt,O.M., R.W. Fujimura, and B.J. Finlayson 1993: Use of <i>Neomysis mercedis</i> (Krebsdyr, Crustacea: Mysidacea) for Estuarine Toxicity Tests. <i>Trans. Am. Fisk, Fish. Soc.</i> 122(2): 279-288
Budischak,S.A., L.K. Belden, and W.A. Hopkins 2009: Relative Toxicity of Malathion to Trematode-Infected and Noninfected <i>Rana palustris</i> Tadpoles. <i>Arch. Environ. Contam. Toxicol.</i> 56(1): 123-128
Butler,P.A. 1963: Commercial Fisk, Fisheries Investigations. Fisk, Fish and Wildlife Service, Circular 167, Washington, D.C.:11-25
Butler,P.A. 1964: Commercial Fisk, Fishery Investigations. In: Pesticide-Wildlife Studies, 1963, U.S.D.I., Fisk, Fish and Wildl.Serv.,Circ.199:28 p
Caldwell,R.S. 1977: Biological Effects of Pesticides on the Dungeness Crab. EPA-600/3-77-131, U.S.EPA, Gulf Breeze, FL:143 p.
Calleja,M.C., G. Persoone, and P. Geladi 1994: Comparative Acute Toxicity of the First 50 Multicentre Evaluation of In Vitro Cytotoxicity Chemicals to Aquatic Non-vertebrates. <i>Arch. Environ. Contam. Toxicol.</i> 26(1): 69-78
Cano,E., A. Jimenez, J.A. Cabral, and M.E. Ocete 1999: Acute Toxicity of Malathion and the New Surfactant "Genapol OXD 080" on Species of Rice Basins. <i>Bull. Environ. Contam.</i>

Toxicol.63(1): 133-138
Carlson,C.A. 1966: Effects of Three Organophosphorus Insecticides on Immature Hexagenia and Hydropsyche of the Upper Mississippi River. Trans. Am. Fisk, Fish. Soc.95(1): 1-5
Chandran,G.J., K.S. Harilal, and S. Sahai 1991: A Method for the Estimation of Safe Experimental Concentration for Intermediate Toxicity Test. J. Nat. Conserv.3(2): 211-217
Chitra,S., and M.K.K. Pillai 1984: Development of Organophosphorus and Carbamate-Resistance in Indian Strains of Anopheles stephensi Liston. Proc. Indian Acad. Sci. Anim. Sci.93(3): 159-170
Cilek,J.E., G.B.,Jr. Craig, and F.W. Knapp 1995: Comparative Susceptibility of Larvae of Three Aedes Species to Malathion and Permethrin. Am. Mosq. Control Assoc. J.11(4): 416-418
Cook,L.W., C.J. Paradise, and B. Lom 2005: The Pesticide Malathion Reduces Survival and Growth in Developing ZebraFisk, Fish. Environ. Toxicol. Chem.24(7): 1745-1750
Cooke, J. 2019: Malathion: Kinetic assessment of laboratory water/sediment studies. Document FMC-53628 in Document M-CA, section 7, Registration documents to EFSA from FMC Agricultural Solutions A/S
Cothran,R.D., R.L. Greco, and R.A. Relyea 2009: No Evidence that a Common Pesticide Impairs Female Mate Choice in a Freshwater Amphipod. Environ. Toxicol.25(3): 310-314
Crisinel,A., L. Delaunay, D. Rossel, J. Tarradellas, H. Meyer, H. Saiah, P. Vogel, C. Delisle, and C. Blaise 1994: Cyst-Based Ecotoxicological Tests Using Anostracans: Comparison of Two Species of Streptocephalus. Environ. Toxicol. Water Qual.9(4): 317-326
Das,P.K., and P.K. Rajagopalan 1979: Susceptibility of Larvae of Culex fatigans (Wiedmann), Anopheles stephensi (Liston) and Aedes aegypti (Linn.) to Insecticides in Pondicherry. Indian J. Med. Res.70:412-416
Ding, T.-T.; Y. Zhang, Y. Zhu, S. Du, J. Zhang, Y. Cao, Y. Wang, G. Wang & L. He 2019: Deriving water quality criteria for China for the organophosphorus pesticides dichlorvos and malathion. Environmental Science and Pollution Research 26: 34622-34632
Dortland (1980): Toxicological evaluation of parathion and azinphosmethyl in freshwater model ecosystems. Agric. Res. Rep. (Versl. landbouwk. Onderz.) 898, ISBN 90 220 0732 4, Doctoral thesis, Wageningen.
EFSA 2009: CONCLUSION ON PESTICIDE PEER REVIEW. Peer review of the pesticide risk assessment of the active substance malathion. EFSA Scientific Report 333, 1-118
FMC 2019
Eisler,R. 1969: Acute Toxicities of Insecticides to Marine Decapod Krebsdyr, Crustaceans. Krebsdyr, Crustaceana (Leiden)16(3): 302-310
Eisler,R. 1970: Acute Toxicities of Organochlorine and Organophosphorus Insecticides to Estuarine Fisk, Fishes. Tech.Pap.No.46, Bur.Sport Fisk, Fish.Wildl., U.S.D.I., Washington, DC:12
EU 2000: Europa-Parlamentets og Rådets Direktiv 2000/60/EF om fastsættelse af en ramme for fællesskabets vandpolitiske foranstaltninger af 23. oktober 2000.
<a href="https://echa.europa.eu/documents/10162/13632/information_requirements_r10_en.pdf/bb902be7-a503-4ab7-9036-d866b8ddce69">EU 2008: ECHA: Guidance on information requirements and chemical safety assessment. Chapter R.10: Characterisation of dose [concentration]-response for environment (https://echa.europa.eu/documents/10162/13632/information_requirements_r10_en.pdf/bb902be7-a503-4ab7-9036-d866b8ddce69)</a>

<p><a href="https://circabc.europa.eu/sd/a/ba6810cd-e611-4f72-9902-f0d8867a2a6b/Guidance%20No%2027%20-%20Deriving%20Environmental%20Quality%20Standards%20-%20version%202018.pdf">EU 2018a: Common Implementation Strategy for the Water Framework Directive (2000/60/EC). Guidance Document No. 27. Technical Guidance for Deriving Environmental Quality Standards. (https://circabc.europa.eu/sd/a/ba6810cd-e611-4f72-9902-f0d8867a2a6b/Guidance%20No%2027%20-%20Deriving%20Environmental%20Quality%20Standards%20-%20version%202018.pdf)</a></p>
<p><a href="#">EU 2018b: European Commission: Final Review report for the active substance malathion finalised in the Standing Committee on the Food Chain and Animal Health at its meeting on 27 November 2009 in view of the inclusion of malathion in Annex I of Directive 91/414/EEC, SANCO/10668/2009 final rev 2, 14 June 2018. (Malathion_Final_Review_Report.pdf)</a></p>
<p>Failloux,A.B., A. Ung, M. Raymond, and N. Pasteur 1994: Insecticide Susceptibility in Mosquitoes (Diptera: Culicidae) from French Polynesia. <i>J. Med. Entomol.</i>31(5): 639-644</p>
<p>Federle,P.F., and W.J. Collins 1976: Insecticide Toxicity to Three Insects from Ohio Ponds. <i>Ohio J. Sci.</i>76(1): 19-24</p>
<p>Fernandez-Casalderry,A., M.D. Ferrando, and E. Andreu-Moliner 1992: Acute Toxicity of Several Pesticides to Rotifer (Brachionus calyciflorus). <i>Bull. Environ. Contam. Toxicol.</i>48(1): 14-17</p>
<p>FMC 2019: Document M-CA Sections 5, 7 and 8 from FMC Agricultural Solutions A/S. Registration documents to EFSA from FMC Agricultural Solutions A/S Indsendt fra konsulentfirmaet FMC Agricultural Solutions A/S til EFSA.</p>
<p>Fordham,C.L., J.D. Tessari, H.S. Ramsdell, and T.J. Keefe 2001: Effects of Malathion on Survival, Growth, Development, and Equilibrium Posture of Bullfrog Tadpoles (<i>Rana catesbeiana</i>). <i>Environ. Toxicol. Chem.</i>20(1): 179-184</p>
<p>Forget,J., J.F. Pavillon, M.R. Menasria, and G. Bocquene 1998: Mortality and LC50 Values for Several Stages of the Marine Copepod <i>Tigriopus brevicornis</i> (Muller) Exposed to the Metals Arsenic and Cadmium and the Pesticides Atrazine, Carbofuran, Dichlorvos, and Malathion. <i>Ecotoxicol. Environ. Saf.</i>40(3): 239-244</p>
<p>Frumin,G.T., G.M. Chuiko, D.F. Pavlov, and O.V. Menzykova 1992: New Rapid Method to Evaluate the Median Effect Concentrations of Xenobiotics in Hydrobionts. <i>Bull. Environ. Contam. Toxicol.</i>49(3): 361-367</p>
<p>Fujimura,R., B. Finlayson, and G. Chapman 1991: Evaluation of Acute and Chronic Toxicity Tests with Larval Striped Bass. <i>ASTM Spec. Tech. Publ.</i>14:193-211</p>
<p>Garza-Leon,C.V., M.A. Arzate-Cardenas, and R. Rico-Martinez 2017. Toxicity Evaluation of Cypermethrin, Glyphosate, and Malathion, on Two Indigenous Zooplanktonic Species. <i>Environ. Sci. Pollut. Res.</i>24:18123-18134</p>
<p>Gaufin,A.R., L.D. Jensen, A.V. Nebeker, T. Nelson, and R.W. Teel 1965: Toxicity of Ten Organic Insecticides to Various Aquatic Invertebrates. <i>Water Sewage Works</i>12:276-279</p>
<p>Giri,A., S.S. Yadav, S. Giri, and G.D. Sharma 2012: Effect of Predator Stress and Malathion on Tadpoles of Indian Skittering Frog. <i>Aquat. Toxicol.</i>106:157-163</p>
<p>Goodman,L.R., G.M. Cripe, P.H. Moody, and D.G. Halsell 1988: Acute Toxicity of Malathion, Tetrabromobisphenol-A, and Tributyltin Chloride to Mysids (<i>Mysidopsis bahia</i>) of Three Ages. <i>Bull. Environ. Contam. Toxicol.</i>41(5): 746-753.</p>
<p>Gul,S., G. Nur, T.O. Kaya, U. Kamber, and B. Gurdegin 2007: Detection of Micronuclei in Peripheral Erythrocytes of <i>Orthrias angorae</i> (Steindachner, 1897) Exposed to Malathion. <i>Fresenius Environ. Bull.</i>16(5): 472-476</p>
<p>Gupta,A.K., D. Dutt, M. Anand, and R.C. Dalela 1994: Combined Toxicity of Chlordane, Malathion and Furadan to a Test Fisk, Fish <i>Notopterus notopterus</i> (Mor). <i>J. Environ. Biol.</i>15(1): 1-6</p>

Guylas,P., and B. Csanyi 1984: Acute Toxic Effect of Different Insecticides on Three Fisk, Fish Species. <i>Aquac. Hung. (Szarvas)</i> 4:79-85
Guzzella,L., A. Gronda, and L. Colombo 1997: Acute Toxicity of Organophosphorus Insecticides to Marine Invertebrates. <i>Bull. Environ. Contam. Toxicol.</i> 59(2): 313-320
Gaaboub,I.A., F.M. El-Gayar, and E.M. Helal 1½975: Comparative Bioassay Studies on Larvae of <i>Culex pipiens</i> and the MicroKrebsdyr, Crustacean <i>Daphnia magna</i> . <i>Bull. Entomol. Soc. Egypt</i> 9:77-84
Haider,S., and R.M. Inbaraj 1986: Relative Toxicity of Technical Material and Commercial Formulation of Malathion and Endosulfan to a Freshwater Fisk, Fish, <i>Channa punctatus</i> (Bloch). <i>Ecotoxicol. Environ. Saf.</i> 11(3): 347-351
Hamed,M.S., A. Ramzi, and S. El Said 1983: Susceptibility Status of Mosquitoes in Egypt to Commonly Used Insectides. <i>J. Egypt. Publ. Health Assoc.</i> 58(1-2): 160-167
Hansen,D.J.. Unpublished Data on the Consent Decree Chemicals. February 22 Memo to W.A.Brungs, U.S.EPA, Gulf Breeze, FL:81 p.
Hansen,D.J., and P.R. Parrish 1977: Suitability of Sheepshead Minnows ( <i>Cyprinodon variegatus</i> ) for Life-Cycle Toxicity Tests. <i>ASTM Spec. Tech. Publ.</i> :117-126
Hermanutz,R.O. 1978: Endrin and Malathion Toxicity to FlagFisk, Fish ( <i>Jordanella floridae</i> ). <i>Arch. Environ. Contam. Toxicol.</i> 7(2): 159-168, 1978.
Hoffman,E.R., and S.W. Fisk, Fisher 1994. Comparison of a Field and Laboratory-Derived Population of <i>Chironomus riparius</i> (Diptera: Chironomidae): Biochemical and Fitness Evidence for Population Divergence. <i>J. Econ. Entomol.</i> 87(2): 318-325
Hoque,M.M., Z. Nahar, and M.A. Hossain 2000: Toxicity of Malathion to Silver Barb ( <i>Barbodes gonionotus</i> Bleeker) Fingerlings. <i>Bangladesh J. Fisk, Fish. Res.</i> 4(1): 101-104
Hua,J., N. Buss, J. Kim, S.A. Orlofske, and J.T. Hoverman 2016. Population-Specific Toxicity of Six Insecticides to the Trematode <i>Echinoparyphium</i> sp.. <i>Parasitology</i> 143:542-550
Jacob,S.S., N.B. Nair, and N.K. Balasubramanian 1982: Toxicity of Certain Pesticides Found in the Habitat to the Larvivorous Fisk, Fishes <i>Aplocheilus lineatus</i> (Cuv. & Val.) and <i>Macropodus cupanus</i> (Cuv. & Val.). <i>Proc. Indian Acad. Sci. Anim. Sci.</i> 91(3): 323-328
Jensen,L.D., and A.R. Gaufin 1964a: Effects of Ten Organic Insecticides on two Species of Stonefly Naiads. <i>Trans. Amer. Fisk, Fish. Soc.</i> , 93(1): 27-34
Jensen,L.D., and A.R. Gaufin 1964b: Long-Term Effects of Organic Insecticides on Two Species of Stonefly Naiads. <i>Trans. Amer. Fisk, Fish. Soc.</i> 93(4): 357-363
Jeyasingam,D.N.T., B. Thayumanavan, and S. Krishnaswamy 1978: The Relative Toxicities of Insecticides on Aquatic Insect <i>Eretes sticticus</i> (Linn.) (Coleoptera: Dytiscidae). <i>J. Madurai Univ.</i> 7(1): 85-87
Katz,M. 1961: Acute Toxicity of Some Organic Insecticides to Three Species of Salmonids and to the Threespine Stickleback. <i>Trans. Am. Fisk, Fish. Soc.</i> 90(3): 264-268.
Kawada,H., Y. Shono, T. Ito, and Y. Abe 1993. Laboratory Evaluation of Insect Growth Regulators Against Several Species of Anopheline Mosquitoes. <i>Med. Entomol. Zool.</i> 44(4): 349-353
Keenleyside,M.H.A. 1958: Comparative Effects of the Insecticides DDT and Malathion on Young Atlantic Salmon. <i>Fisk, Fish. Res. Board Can. Atl. Reg. Off. Circ.</i> 69:3-6
Keller,A.E., and D.S. Ruessler 1997: The Toxicity of Malathion to Unionid Mussels: Relationship to Expected Environmental Concentrations. <i>Environ. Toxicol. Chem.</i> 16(5): 1028-1033
Key,P.B. 1995: The Lethal and Sublethal Effects of Malathion, Azinphosmethyl and Chlorpyrifos Exposure on the Grass Shrimp, <i>Palaemonetes pugio</i> , with Emphasis on Larval Life Cycle Pulse Exposures. Ph.D.Thesis, University of South Carolina, Columbia, SC:124 p

Key,P.B., M.H. Fulton, G.I. Scott, S.L. Layman, and E.F. Wirth 1998. Lethal and Sublethal Effects of Malathion on Three Life Stages of the Grass Shrimp, <i>Palaemonetes pugio</i> . <i>Aquat. Toxicol.</i> 40(4): 311-322
Key,P.B., and M.H. Fulton 2006: Correlation Between 96-h Mortality and 24-h Acetylcholinesterase Inhibition in Three Grass Shrimp Larval Life Stages. <i>Ecotoxicol. Environ. Saf.</i> 63(3): 389-392
Khangarot,B.S., A. Sehgal, and M.K. Bhasin 1985: "Man and Biosphere" - Studies on the Sikkim Himalayas. Part 6: Toxicity of Selected Pesticides to Frog Tadpole <i>Rana hexadactyla</i> (Lesson). <i>Acta Hydrochim. Hydrobiol.</i> 13(3): 391-394
Khangarot,B.S., and P.K. Ray 1988: Studies on the Toxicity of Malathion to Freshwater Teleosts, <i>Channa punctatus</i> (Bloch) and <i>Puntius sophore</i> (Hamilton). <i>Arch. Hydrobiol.</i> 113(3): 465-469.
Khattak,I.U.D., and M.A. Hafeez 1996: Effect of Malathion on Blood Parameters of the Fisk, Fish, <i>Cyprinion watsoni</i> . <i>Pak. J. Zool.</i> 28(1): 45-49
Khillare,Y.K., and S.B. Wagh 1988: Long-Term Effects of Pesticides Endosulfan, Malathion and Sevin on the Fisk, Fish <i>Puntius stigma</i> . <i>Environ. Ecol.</i> 6(3): 589-593
Kline,D.L., J.R. Wood, R.H. Roberts, and K.F. Baldwin 1985: Laboratory Evaluation of Four Organophosphate Compounds As Larvicides Against Field Collected Salt Marsh <i>Culicoides</i> spp. (Diptera: Ceratopogonidae). <i>Am. Mosq. Control Assoc. J.</i> 1(1): 48-50
Knoch, E. 2001: Degradability and fate of malathion in the aquatic environment (water/sediment system). Document 337-FYF in Document M-CA, section 7, Registration documents to EFSA from FMC Agricultural Solutions A/S
Komalamisra,N., Y. Trongtokit, K. Palakul, S. Prummongkol, Y. Samung, C. Apiwathnasorn, T. Phanpoowong, A. Asavanich 2006 a. Insecticide Susceptibility of Mosquitoes Invading Tsunami-Affected Areas of Thailand. <i>Southeast Asian J. Trop. Med. Public Health</i> 37(suppl. 3): 118-122
Konar,S.K., and T.K. Ghosh 1981: Effects of Organophosphorus Insecticides on Fisk, Fish and Fisk, Fish Food Organisms. <i>IAWPC Tech.Annu.</i> 8:147-160
Kumari,S.D.R., and N.B. Nair 1978: Toxicity of Some Insecticides to <i>Lepidocephalus thermalis</i> (Cuv. & Val.). <i>Proc. Natl. Acad. Sci. India Sect. B Biol. Sci.</i> 44(3): 122-132
Kusk,K.O., A.M. Christensen, and N. Nyholm2018. Algal Growth Inhibition Test Results of 425 Organic Chemical Substances. <i>Chemosphere</i> 204:405-412
Lahr,J., A. Badji, S. Marquenie, E. Schuiling, K.B. Ndour, A.O. Diallo, and J.W. Everts 2001: Acute Toxicity of Locust Insecticides to Two Indigenous Invertebrates from Sahelian Temporary Ponds. <i>Ecotoxicol. Environ. Saf.</i> 48(1): 66-75
Leight,A.K., and R.F. Van Dolah 1999:. Acute Toxicity of the Insecticides Endosulfan, Chlorpyrifos, and Malathion to the Epibenthic Estuarine Amphipod <i>Gammarus palustris</i> (Bousfield). <i>Environ. Toxicol. Chem.</i> 18(5): 958-964
Lewis,W.M. 1993. Acute and Chronic Responses of <i>Menidia beryllina</i> , <i>Rivulus marmoratus</i> , and <i>Cyprinodon variegatus</i> to Malathion and Zinc Chloride. M.S.Thesis, Florida Institute of Technology, Melbourne, FL:59 p.
Li,S.N., and D.F. Fan 1996: Correlation Between Biochemical Parameters and Susceptibility of Freshwater Fisk, Fish to Malathion. <i>J. Toxicol. Environ. Health</i> 48(4): 413-418, 1996. ECOREF #19981
Liong,P.C., W.P. Hamzah, and V. Murugan 1988: Toxicity of Some Pesticides Towards Freshwater Fisk, Fishes. <i>Fisk, Fish.Bull.Dep.Fisk, Fish.(Malays.)</i> No.57:13 p.
Liu,D.H.W., and J.M. Lee 1975: Toxicity of Selected Pesticides to the Bay Mussel ( <i>Mytilus edulis</i> ). EPA-660/3-75-016, U.S.EPA, Corvallis, OR:102 p
Lomte,V.S., and S.M. Alam 1985: Temperature Dependent Toxicity of Malathion to the

Freshwater Gastropod <i>Viviparus bengalensis</i> . <i>Geobios</i> (Jodhpur)12(6): 251-253
Mane,U.H., S.R. Akarte, and D.V. Muley 1984: Effect of Cythion-Malathion on Respiration in Three Freshwater Bivalve Molluscs From Godavari River Near Paithan. <i>J. Environ. Biol.</i> 5(2): 71-80
Mane,U.H., and D.V. Muley 1987: Seasonal Variations in the Toxicity of Cythion-Malathion to Two Freshwater Bivalve Molluscs. <i>Comp. Physiol. Ecol.</i> 12(1): 25-31
Mayer,F.L.,Jr., and M.R. Ellersieck 1986. Manual of Acute Toxicity: Interpretation and Data Base for 410 Chemicals and 66 Species of Freshwater Animals. USDI Fisk, Fish and Wildlife Service, Publication No.160, Washington, DC:505 p.
Milam,C.D., J.L. Farris, and J.D. Wilhide 2000: Evaluating Mosquito Control Pesticides for Effect on Target and Nontarget Organisms. <i>Arch. Environ. Contam. Toxicol.</i> 39(3): 324-328
Miljøstyrelsen 2004: Principper for fastsættelse af vandkvalitetskriterier for stoffer i overfladevand. Vejledning fra Miljøstyrelsen nr. 4, 2004.
Moermond, C.T., Kase, R., Korkaric, M. and Ågerstrand, M. (2016). CRED: Criteria for reporting and evaluating ecotoxicity data. <i>Environ Toxicol Chem</i> , 35, s. 1297-1309.
Mohsin,A.K.M., O.S. Lui, and M.A. Ambak 1984. Effect of Malathion on Sepat Siam and Tilapia. <i>Pertanika</i> 7(1): 57-60
Montada,D., R. Tang, A. Navarro, and F.A. Garcia 1994: Susceptibility Status of <i>Aedes taeniorhynchus</i> to Organochlorine and Organophosphate Insecticides. <i>Mem. Inst. Oswaldo Cruz</i> 89(2): 251-252
Mulla,M.S., R.L. Metcalf, and G. Kats 1964. Evaluation of New Mosquito Larvicides, with Notes on Resistant Strains. <i>Mosq. News</i> 24(3): 312-319
Nagabhushanam,R., G. Gyananath, and R. Sarojini 1983: Toxicity of Three Organophosphate Compounds and Their Effect on the Gonad of Freshwater Prawn, <i>Macrobrachium lamerrii</i> . <i>Indian J. Comp. Anim. Physiol.</i> 1(1): 71-75
Nalecz-Jawecki,G., E. Kucharczyk, and J. Sawicki 2002: The Sensitivity of Protozoan <i>Spirostomum ambiguum</i> to Selected Pesticides. <i>Fresenius Environ. Bull.</i> 11(2): 98-101
Natarajan,E., R.S. Biradar, and J.P. George 1992: Acute Toxicity of Pesticides to Giant Freshwater Prawn <i>Macrobrachium rosenbergii</i> (De Man). <i>J. Aquacult. Trop.</i> 7(2): 183-187
Nishiuchi,Y., and K. Asano 1978: Toxicity of Formulated Agrochemicals to Fresh Water Organisms LII. <i>Suisan Zoshoku</i> 26(1): 26-30
Nishiuchi,Y., and Y. Hashimoto 1969: Toxicity of Pesticides to Some Fresh Water Organisms. <i>Rev. Plant Prot. Res.</i> 2:137-139
Nogrady,T., and J. Keshmirian 1986: Rotifer Neuropharmacology - I. Cholinergic Drug Effects on Oviposition of <i>Philodina acuticornis</i> (Hjuldyr, Rotifera, Aschelminthes). <i>Comp. Biochem. Physiol. C Comp. Pharmacol.</i> 83(2): 335-338
Ocampo,C.B., W.G. Brogdon, C.M. Orrego, G. Toro, and J. Montoya-Lerma 2000: Insecticide Susceptibility in <i>Anopheles pseudopunctipennis</i> from Colombia: Comparison Between Bioassays and Biochemical Assays. <i>Am. Mosq. Control Assoc. J.</i> 16(4): 331-338
Olvera-Hernandez,E., L. Martinez-Tabche, and F. Martinez-Jeronimo 2004: Bioavailability and Effects of Malathion in Artificial Sediments on <i>Simocephalus vetulus</i> (Cladocera: Daphniidae). <i>Bull. Environ. Contam. Toxicol.</i> 73(1): 197-204
Overmyer,J.P., K.L. Armbrust, and R. Noblet 2003: Susceptibility of Black Fly Larvae (Diptera: Simuliidae) to Lawn-Care Insecticides Individually and as Mixtures. <i>Environ. Toxicol. Chem.</i> 22(7): 1582-1588
Pan,D.Y., and X.M. Liang 1993: Safety Study of Pesticides on Bog Frog, a Predatory Natural Enemy of Pest in Paddy Field. <i>J. Hunan Agric. Coll.</i> 19(1): 47-54

Patil,V.K., and M. David 2010: Behavioral and Morphological Endpoints: As an Early Response to Sublethal Malathion Intoxication in the Freshwater Fisk, Fish, <i>Labeo rohita</i> . <i>Drug Chem. Toxicol.</i> 33(2): 160-165
Pickering,Q.H., C. Henderson, and A.E. Lemke 1962: The Toxicity of Organic Phosphorus Insecticides to Different Species of Warmwater Fisk, Fishes. <i>Trans. Am. Fisk, Fish. Soc.</i> 91:175-184
Piri,M., and V. Ordog 1999: Herbicides and Insecticides Effects on Green Algae and Cyanobacteria Strain. <i>Iran. J. Fisk, Fish. Sci.</i> 1(1): 47-58
Ponlawat,A., J.G. Scott, and L.C. Harrington 2005: Insecticide Susceptibility of <i>Aedes aegypti</i> and <i>Aedes albopictus</i> Across Thailand. <i>J. Med. Entomol.</i> 42(5): 821-825
Post,G., and T.R. Schroeder 1971: The Toxicity of Four Insecticides to Four Salmonid Species. <i>Bull. Environ. Contam. Toxicol.</i> 6(2): 144-155
Rao,T.S., S. Dutt, and K. Mangaiah 1967: TLM Values of Some Modern Pesticides to the Freshwater Fisk, Fish - <i>Puntius puckelli</i> . <i>Environ. Health (London)</i> 9:103-109, 1967. ECOREF #6722
Rao,M.B., and U.K. Mane 1978: The Effect of Carbofos on the Survival Rate and Respiration of Black Sea Mussels ( <i>Mytilus galloprovincialis</i> ). <i>Hydrobiol. J.</i> 14(6): 90-94
Rao,K.S., and R. Nagabhushanam 1987: Toxicity of DDT and Malathion to the Freshwater Crab <i>Barytelphusa cunicularis</i> . <i>Aquat. Sci. Fisk, Fish. Abstr.</i> 17(8, Pt. 1): 11842-1Q17, 1987. ECOREF #12790
Reddy,M.S., and K.V.R. Rao 1992: Toxicity of Selected Insecticides to the Penaeid Prawn, <i>Metapenaeus monoceros</i> (Fabricius). <i>Bull. Environ. Contam. Toxicol.</i> 48(4): 622-629
Rehwoldt,R.E., E. Kelley, and M. Mahoney 1977: Investigations into the Acute Toxicity and Some Chronic Effects of Selected Herbicides and Pesticides on Several Fresh Water Fisk, Fish Species. <i>Bull. Environ. Contam. Toxicol.</i> 18(3): 361-365
Reid,H.P., J.T. Ahokas, and D.A. Holdway 1995: Use of Cyanazine and Malathion Pulse-Exposure Toxicity to Estimate the Age of Onset of Functional Liver Metabolism in Larval Australian Crimson-Spotted RainbowFisk, Fish ( <i>Melanotaenia fluviatilis</i> ). <i>Water Res.</i> 29(8): 2010-2013
Relyea,R.A. 2004: Synergistic Impacts of Malathion and Predatory Stress on Six Species of North American Tadpoles. <i>Environ. Toxicol. Chem.</i> 23(4): 1080-1084
Rettich,F. 1977: The Susceptibility of Mosquito Larvae to Eighteen Insecticides in Czechoslovakia. <i>Mosq. News</i> 37(2): 252-257
Reyes,J.G.G., N.R. Leyva, O.A. Millan, and G.A. Lazcano 2002: Effects of Pesticides on DNA and Protein of Shrimp Larvae <i>Litopenaeus stylirostris</i> of the California Gulf. <i>Ecotoxicol. Environ. Saf.</i> 53(2): 191-195
Ripley,B.J., K.C. Davis, B.J. Carter, and M.A. Simovich 2002: Toxicity of Malathion and Roundup to the San Diego Fairy Shrimp. <i>Trans. West. Wildl. Soc.</i> 38/39:13-21
Rubio-Moran,R., W.A. Rowley, and J.R. Coats 1981: Effect of Permethrin and Malathion on <i>Aedes trivittatus</i> and <i>Culex pipiens pipiens</i> . <i>Iowa State J. Res.</i> 55(3): 235-244
Sailatha,D., I.K.A. Sahib, and K.V.R. Rao 1981: Toxicity of Technical and Commercial Grade Malathion to the Fisk, Fish, <i>Tilapia mossambica</i> (Peters). <i>Proc. Indian Acad. Sci. Anim. Sci.</i> 90(1): 87-92
Salame 1996:
Sanders,H.O. 1972: Toxicity of Some Insecticides to Four Species of Malacostracan Krebsdyr, Crustaceans. <i>Tech. Pap. Bur. Sport Fisk, Fish. Wildl.</i> 66:19 p.
Sayim,F. 2008. Acute Toxic Effects of Malathion on the 21st Stage Larvae of the Marsh Frog.



Turk. J. Zool.32:99-106
Schoettger,R.A. 1970: Fisk, Fish-Pesticide Research Laboratory. In: Prog. in Sport Fisk, Fish. Res., U.S. Dep. Interior, Bur. Sport Fisk, Fish. and Wildl. Res., Publ. No.106:2-40
Scirocchi,A., and A. D'Erme 1980: Toxicity of Seven Insecticides on Some Species of Fresh Water Fisk, Fishes. Riv. Parassitol.41(1): 113-121
Scott,G.I., D.S. Baughman, A.H. Trim, and J.C. Dee 1987: Lethal and Sublethal Effects of Insecticides Commonly Found in Nonpoint Source Agricultural Runoff to Estuarine Fisk, Fish and ShellFisk, Fish. In: W.B.Vernberg, A.Calabrese, F.P.Thurberg, and F.J.Vernberg (Eds.), Pollution Physiology of Estuarine Organisms, University of South Carolina Press, Columbia, SC:251-273
Shedd,T.R., M.W. Widder, M.W. Toussaint, M.C. Sunkel, and E. Hull 1999. Evaluation of the Annual KilliFisk, Fish <i>Nothobranchius guentheri</i> as a Tool for Rapid Acute Toxicity Screening. Environ. Toxicol. Chem.18(10): 2258-2261
Shim,J.C., and L.S. Self 1973: Toxicity of Agricultural Chemicals to Larvivorous Fisk, Fish in Korean Rice Fields. Trop. Med.15(3): 123-130
Singh,S., and S. Sahai 1984. Toxicity of Some Pesticide to Two Fresh Water Teleosts. J. Environ. Biol.5(4): 255-259
Sinha,N., B. Lal, and T.P. Singh 1991: Pesticides Induced Changes in Circulating Thyroid Hormones in the Freshwater CatFisk, Fish <i>Clarias batrachus</i> . Comp. Biochem. Physiol. C Comp. Pharmacol.100(1/2): 107-110
Snawder,J.E., and J.E. Chambers 1989: Toxic and Developmental Effects of Organophosphorus Insecticides in Embryos of the South African Clawed Frog. J. Environ. Sci. Health Part B Pestic. Food Contam. Agric. Wastes24(3): 205-218
Snell,T.W., and G. Persoone 1989: Acute Toxicity Bioassays Using Rotifers. II. A Freshwater Test with <i>Brachionus rubens</i> . Aquat. Toxicol.14(1): 81-92.
Sparling,D.W., and G. Fellers 2007: Comparative Toxicity of Chlorpyrifos, Diazinon, Malathion and Their Oxon Derivatives to Larval <i>Rana boylii</i> . Environ. Pollut.147(3): 535-539
Sreenivasan,A., and G.K. Swaminathan 1967: Toxicity of Six Organophosphorus Insecticides to Fisk, Fish. Curr. Sci. (Bangalore)36(15): 397-398
Stevens,M.M. 1992: Toxicity of Organophosphorus Insecticides to Fourth-Instar Larvae of <i>Chironomus tepperi</i> Skuse (Diptera: Chironomidae). J. Aust. Entomol. Soc.31:335-337
Tchounwou,P.B., A.J.,Jr. Englande, E.A. Malek, G.J. Greer, and A.C. Anderson 1991: The Effects of Bayluscide and Malathion on the Mortality and Infectivity of <i>Schistosoma mansoni</i> Cercariae. Environ. Toxicol. Water Qual.7(2): 107-117
Tchounwou,P.B., A.J.,Jr. Englande, E.A. Malek, G.J. Greer, and A.C. Anderson 1992: The Effects of Bayluscide and Malathion on the Mortality and Infectivity of <i>Schistosoma mansoni</i> Cercariae. Environ. Toxicol. Water Qual.7(2): 107-117
Tietze,N.S., M.A. Olson, P.G. Hester, and J.J. Moore 1993: Tolerance of Sewage Treatment Plant Microorganisms to Mosquitocides. Am. Mosq. Control Assoc. J.9(4): 477-479
Toumi, H; M. Boumaiza, M. Millet, C.M. Radetski, B.I. Camara, V. Felten, J. Ferard 2015: Investigation of differences in sensitivity between 3 strains of <i>Daphnia magna</i> (Krebsdyr, Crustacean Cladocera) exposed to malathion (organophosphorous pesticide). Journal of Environmental Science and Health, Part B: Pesticides, Food Contaminants, and Agricultural Wastes (2015), 50(1), 34-44
Tsuda,T., M. Kojima, H. Harada, A. Nakajima, and S. Aoki 1997: Acute Toxicity, Accumulation and Excretion of Organophosphorous Insecticides and Their Oxidation Products in KilliFisk,

Fish. Chemosphere35(5): 939-949
U.S. Environmental Protection Agency. Pesticide Ecotoxicity Database (Formerly: Environmental Effects Database (EEDB)). Environmental Fate and Effects Division, U.S.EPA, Washington, D.C., 1992
Varanka,I. 1986: Toxicity of Mosquitocides on Freshwater Mussel Larvae. Acta Biol. Hung.37(2): 143-158
Varanka et Dhande 1991:
Vedamanikam,V.J. 2009: Formation of Resistance in the Chironomus plumosus to Four Pesticides over 45 Generations. Toxicol. Environ. Chem.91(1): 187-194
Venturino,A., L.E. Gauna, R.M. Bergoc, and A.M.P. D'Angelo 1992: Effect of Exogenously Applied Polyamines on Malathion Toxicity in the Toad Bufo arenarum Hensel. Arch. Environ. Contam. Toxicol.22:135-139
Verep,B. 2006: Toxicity of Malathion on European Chub. Indian Vet. J.83(8): 912-913
Verma,S.R., I.P. Tonk, and R.C. Dalela 1981: Determination of the Maximum Acceptable Toxicant Concentration (MATC) and the Safe Concentration for Certain Aquatic Pollutants. Acta Hydrochim. Hydrobiol.9(3): 247-254
Verma,S.R., S.K. Bansal, A.K. Gupta, N. Pal, A.K. Tyagi, M.C. Bhatnagar, V. Kumar, and R.C. Dalela 1982: Bioassay Trials with Twenty-Three Pesticides to a Fresh Water Teleost, Saccobranchus fossilis. Water Res.16(5): 525-529
Verma,S.R., I.P. Tonk, A.K. Gupta, and M. Saxena 1984: Evaluation of an Application Factor for Determining the Safe Concentration of Agricultural and Industrial Chemicals. Water Res.18(1): 111-115
Victor,B. 1989: Histological Evaluation of Cythion Toxicity on the Oocyte Development of Oziotelphusa senex senex. J. Environ. Biol.10(1): 65-71
Vijayan,V.A., M.A. Revanna, K.S. Vasudeva, N. Pushpalatha, and Poornima 1993:. Comparative Susceptibility of Two Japanese Encephalitis Vectors from Mysore to Six Insecticides. Indian J. Med. Res. Sect. A97:215-217
Villar,D., M.H. Li, and D.J. Schaeffer 1993: Toxicity of Organophosphorus Pesticides to Dugesia dorotocephala. Bull. Environ. Contam. Toxicol.51:80-87
Villar,D., M. Gonzalez, M.J. Gualda, and D.J. Schaeffer 1994. Effects of Organophosphorus Insecticides on Dugesia tigrina: Cholinesterase Activity and Head Regeneration. Bull. Environ. Contam. Toxicol.52(2): 319-324
Wagner, C. & Løkke, H. 1991: Estimation of ecotoxicological protection levels from NOEC toxicity data. Water Research, <b>25</b> , 1237-1242.
Whitten,B.K., and C.J. Goodnight 1966: Toxicity of Some Common Insecticides to Tubificids. J. Water Pollut. Control Fed.38(2): 227-235
Wilkins,R.M., and R.J. Metcalfe 1993: Toxicity of Soil Applied Herbicides to Brine Shrimp Larvae (Artemia salina) and Synergism with Other Pesticides. Br. Crop Prot. Conf. Weeds1:163-168
Winner,R.A., C.D. Steelman, and P.E. Schilling 1978. Effects of Selected Insecticides on Romanomermis culicivorax, a Mermithid Nematode Parasite of Mosquito Larvae. Mosq. News38(4): 546-552
Yang,T., and F. Zhang 1989: Acute Toxicity of 7 Pesticides on 2 Species of Common Leeches. China Environ. Sci.9(1): 51-55
Yeh,H.J., and C.Y. Chen 2006: Toxicity Assessment of Pesticides to Pseudokirchneriella subcapitata Under Air-Tight Test Environment. J. Hazard. Mater.131(1-3): 6-12
Yeragi,S.G., and V.A. Koli 1999: Effect of Malathion on Respiration of Marine Edible Crab Uca

marionis (Des). J. Mar. Biol. Assoc. India41(1-2): 129-133
Yokoyama,T., H. Saka, S. Fujita, and Y. Nishiuchi 1988: Sensitivity of Japanese Eel, <i>Anguilla japonica</i> , to 68 Kinds of Agricultural Chemicals. Bull. Agric. Chem. Insp. Stn. (Tokyo)28:26-33
Zahran,H.E.D.M., and S.A.M. Abdelgaleil 2011: Insecticidal and Developmental Inhibitory Properties of Monoterpenes on <i>Culex pipiens</i> L. (Diptera: Culicidae). J. Asia Pac. Entomol.14(1): 46-51
Ziegler,R., S. Whyard, A.E.R. Downe, G.R. Wyatt, and V.K. Walker 1987: General Esterase, Malathion Carboxylesterase, and Malathion Resistance in <i>Culex tarsalis</i> . Pestic. Biochem. Physiol.28(2): 279-285
Zulkosky,A.M., J.P. Ruggieri, S.A. Terracciano, B.J. Brownawell, and A.E. McElroy 2005: Acute Toxicity of Resmethrin, Malathion and Methoprene to Larval and Juvenile American Lobsters ( <i>Homarus americanus</i> ) and Analysis of Pesticide Levels in Surface Waters After Scourge, Anvil and Altosid Application. J. ShellFisk, Fish Res.24(3): 795-804

# Bilag A

## Giftighed over for vandorganismer (EC<sub>x</sub>, LC<sub>x</sub>, NOEC, osv.)

### Ferskvandsorganismer

Akut giftighed

*Freshwater, acute toxicity*

Akutte data for malathion, EC<sub>50</sub> og LC<sub>50</sub>. For alger og cyanobakterier bruges normalt vækstraten, dvs. E<sub>r</sub>C<sub>50</sub>, hvis den er tilgængelig. Alle referencer fra FMC (2019) betragtes som troværdige med et troværdighedsindeks på RI1 eller RI2. Da disse studier ofte bare angives som "acceptable" er der i tabellen ikke angivet en RI-værdi udfor FMC-studier.

Acute toxicity data for malathion, EC<sub>50</sub> and LC<sub>50</sub>. For algae and cyanobacteria the EC<sub>50</sub> is normally for growth rate, i.e. E<sub>r</sub>C<sub>50</sub> is employed if available. All references from FMC 2019 are regarded as reliable with a reliability index of RI1 or RI2

Overordnet systematiske gruppe	Art	EC <sub>50</sub> µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
<b>Cyanobacteria</b>	<i>Anabaena flos-aquae</i>	41425	3	y	96		FMC 2019: Dobbins et al. 2012, 1262 FYF	
<b>Alger ( Algae) Chlorophyta</b>	<i>Raphidocelis subcapitata</i>	131218	3	y	96,4		FMC 2019: Jenkins 1993, 071 FYF, FMC 2019: Dobbins 2012, 1246 FYF Yeh & Chen 2006	Geometrisk gennemsnit af de tre studier. Yeh & Chen studiet havde den laveste værdi (2040 µg/l). Værdien er fra ECOTOX og er ikke blevet evalueret. Endvidere var koncentrationerne ikke målt og renheden af stoffet ikke angivet.  Konsekvensen af brugen af nominelle koncentrationer og af at have en lavere renhed vil være, at man overvurderer koncentrationen, dvs. får en større EC <sub>50</sub> end den skulle være.  Da dette studie giver den laveste værdi, vil det være betænkeligt at udelade det.

Overordnet systematiske gruppe	Art	EC <sub>50</sub> µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
	<i>Scenedesmus obtusiusculus</i>	55380	6	n	60		Piri & Ordog 1999	
<b>Bacillariophyceae</b>	<i>Navicula pelliculosa</i>	23669	3	y	96		FMC 2019: Dobbins et al 2012, 1245 FYF,	
<b>Højere planter (Macrophyta)</b>	<i>Lemna gibba</i>	81150	7	y	96		FMC 2019: Dobbins et al 2012, 1264 FYF,	
<b>Protozoa</b>	<i>Spirostomum ambiguum</i>	39100	1	n			Nalecz-Jawecki et al 2002	
<b>Protozoa</b>	<i>Tetrahymena pyriformis</i>	32000	0,375	n	90		Bogaerts et al 2001	Ekspontential vækstfase (log)
<b>Fladorme (Platyhelminthes) Trematoda</b>	<i>Schistosoma mansoni</i>	69360	0,167	n	91		Tchounwou et al 1992	
<b>Fladorme (Platyhelminthes) Turbellaria</b>	<i>Dugesia dorotocephala</i>	6000	7	n			Villar et al 1993	
	<i>Girardia tigrina</i>	4400	4	n	54,4		Villar et al 1994	
<b>Hjuldyr (Rotifera)</b>	<i>Brachionus calyciflorus</i>	33720	1	n	95		Fernandez-Casalderry et al 1992	Cyst
	<i>Brachionus rubens</i>	35300	1	n			Snell et Persoone 1989	Neonat
	<i>Lecane papuana</i>	42840	2	n	97,1		Garza-Leon et al 2017	Neonate
	<b><i>Philodina acuticornis</i></b>	<b>165,176</b>	3	n			Nogrady et Keshmirian 1986	Voksne
<b>Rundorme (Nematoda)</b>	<i>Romanomermis culicivorax</i>	6780	1	n			Winner et al 1978	

Overordnet systematiske gruppe	Art	EC <sub>50</sub> µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
<b>Bløddyr (Mollusca) Bivalvia</b>	<i>Anodonta anatina</i>	2030	2	n	95	2	Varanka 1986	Larve
	<i>Anodonta cygnea</i>	10210	2	n	95	2	Varanka 1986	Larve
	<i>Elliptio icterina</i>	32000	4	n	96	2	Keller et Ruessler 1997	Juvenile
	<i>Indonaia caerulea</i>	<b>12</b>	4	n	95		Mane et al 1984	
	<i>Lamellidens corrianus</i>	<b>118,55</b>	4	n			Mane et Muley 1987	Voksne
	<i>Lamellidens marginalis</i>	<b>55,63</b>	4	n			Mane et Muley 1987	Voksne
	<i>Lampsilis siliquoidea</i>	7000	2	n	96	2	Keller et Ruessler 1997	Voksne
	<i>Lampsilis straminea ssp. claibornensis</i>	24000	4	n	96	2	Keller et Ruessler 1997	Juvenile
	<i>Lampsilis subangulata</i>	28000	4	n	96	2	Keller et Ruessler 1997	Juvenile. Blev faktisk malt, men var tæt på nominelle
	<i>Lampsilis teres</i>	28000	0,167	n	96	2	Keller et Ruessler 1997	Glochidia. Blev faktisk malt, men var tæt på nominelle
	<i>Megalonaias nervosa</i>	22000	1	n	96	2	Keller et Ruessler 1997	Voksne
	<i>Utterbackia imbecillis</i>	215000	4	n	96	2	Keller et Ruessler 1997	Juvenile. Blev faktisk malt, men var tæt på nominelle
	<i>Villosa lienosa</i>	54000	1	n	96	2	Keller et Ruessler 1997	Voksne
	<i>Villosa villosa</i>	117000	1	n	96	2	Keller et Ruessler 1997	Voksne

Overordnet systematiske gruppe	Art	EC <sub>50</sub> µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
<b>Bløddyr (Mollusca) Gastropoda</b>	<i>Bellamyia bengalensis</i>	17500	4	n			Vankhede et Dhande 1991	
	<i>Biomphalaria havanensis</i>	94780	1	n	91	2	Tchounwou et al 1991	Æg
	<i>Helisoma duryi</i>	1760	1	n			Bakry et al 2011	Voksne
	<i>Helisoma trivolvis</i>	187650	1	n	91,5	2	Tchounwou et al 1991	Æg (fostre)
	<b><i>Lymnaea acuminata</i></b>	<b>16,9</b>	2	n	50		Chaudhari et al 1988	
	<i>Lymnaea stagnalis</i>	7400	2	n	40		Frumin et 1992	Voksne
	<i>Pila globosa</i>	10000	2	n	50		Ahamad et al 1978	
	<i>Planorbella trivolvis</i>	187650	1	n	91	2	Tchounwou et al 1991	Æg
	<i>Planorbis corneus</i>	62000	2	n	40		Frumin et 1992	Voksne
	<i>Pomacea canaliculata</i>	18680	2	n			Tejada et al 1994	
	<b><i>Thiara lineata</i></b>	<b>31,1</b>	2	n	50		Chaudhari et al 1988	
	<b><i>Thiara scabra</i></b>	<b>37,4</b>	2	n	50		Chaudhari et al 1988	
	<i>Viviparus bengalensis</i>	1510	4	n			Lomte et Alam 1985	
	<b>Ledorme (Annelida) Oligochaeta</b>	<i>Lumbriculus variegatus</i>	20500	4	n			Bailey et Liu 1980

Overordnet systematiske gruppe	Art	EC <sub>50</sub> µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
	<i>Tubifex sp.</i>	43100	2	n	40		Frumin et al 1992	Voksne
	<i>Tubificidae</i>	20700	2	n	99,6		Whitten et Goodnight 1966	
<b>Ledorme (Annelida)</b> <b>Hirudinea</b>	<i>Erpobdella octoculata</i>	27000	4	n			Yang et Zhang 1989	
	<i>Glossiphonia sp.</i>	13000	4	n			Yang et Zhang 1989	
<b>Krebsdyr (Crustacea)</b> <b>Amphipoda</b>	<i>Gammarus fasciatus</i>	<b>0,48</b>	5	n		2	Sanders 1972	"Flow-through"
	<i>Gammarus lacustris</i>	<b>1,8</b>	2	n			US EPA 1992	II tre andre studier med RI3 og RI4 lå værdierne på 1,6, 1,8 og 3,8 µg/l, altså meget tæt på den her viste værdi på 1,8. Så selvom studierne ikke er troværdige understøtter de samlet set værdien på 1,8 µg/l
	<i>Gammarus pseudolimnaeus</i>	<b>2,1</b>	4	y	96,8		FMC 2019: Brougher et al 2014, 1417 FYF	
	<i>Gammarus pulex</i>	<b>0,33</b>	4	y	97,8	2	Ashauer et al 2011	Voksne
	<i>Hyaella sp.</i>	<b>0,06</b>	4	y	99,1		Cothran et al 2009	
<b>Krebsdyr (Crustacea)</b> <b>Branchiopoda</b>	<i>Alona guttata</i>	<b>5,26</b>	2	n	97,1		Garza-Leon et al 2017	Neonate
	<i>Ceriodaphnia dubia</i>	<b>2,12</b>	2	n			Ankley et al 1991	



Overordnet systematiske gruppe	Art	EC <sub>50</sub> µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
	<i>Daphnia magna</i>	0,27	1	n		2	Gaaboub et al 1975	Neonate
	<i>Daphnia magna</i>	0,72	2	y	96,9		FMC 2019: Gries et Purghart 2201, 310 FYF	
	<i>Daphnia magna</i>	0,75	2	y	96,8		FMC 2019: Juckeland 2019, 4136 ETX-FYF	Lavt isomalathion content
	<i>Daphnia magna</i>	0,8	2	y	96		FMC 2019: Juckeland 2019, 4136 ETX-FYF	Højt isomalathion i indhold. QSAR forudsigelser (danske QSAR database, <a href="https://qsar.db.food.dtu.dk/db/index.html">https://qsar.db.food.dtu.dk/db/index.html</a> ) estimerer en lavere giftighed af isomalathion overfor vandlevende organismer end for malathion.
	<i>Daphnia magna</i>	2,1	2	y	99,4		FMC 2019: Mattock 20021, 369 FYF	
	<i>Daphnia magna</i>	11,32	2	n	95		FMC 2019: Zhang et al 2011	Racemisk blanding
	<i>Daphnia magna</i>	6,94	2	n	95		FMC 2019: Zhang et al 2011	R-malathion
	<i>Daphnia magna</i>	11,62	2	n	95		FMC 2019: Zhang et al 2011	S-malathion
	<i>Daphnia magna</i>	0,36	2	n	50		FMC 2019:Toumi et al 2015	Strain S1
	<i>Daphnia magna</i>	0,44	2	n	50		FMC 2019:Toumi et al 2015	Strain S2
	<i>Daphnia magna</i>	0,46	2	n	50		FMC 2019:Toumi et al 2015	Strain S3
	<b><i>Daphnia magna</i></b>	<b>1,3</b>						Geometrisk gennemsnit af <i>D. magna</i> værdierne ovenfor.
	<i>Daphnia similis</i>	1710	2	y	1113 g/l		FMC 2019: Perina et al 1995, 248 FYF	
	<b><i>Moina macrocopa</i></b>	<b>58</b>	0,125	n			Nishiuchi et Hashimoto 1969	Voksne

Overordnet systematiske gruppe	Art	EC <sub>50</sub> µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
	<b><i>Simocephalus vetulus</i></b>	<b>2,9</b>	2	y			Olvera-Hernandez et al 2004	Neonate
<b>Krebsdyr (Crustacea) Branchiopoda Anostraca</b>	<i>Branchinecta sandiegonensis</i>	24450	1	n			Ripley et al 2002	
	<i>Streptocephalus proboscideus</i>	64418,45	1	n			Calleja et al 1994	
	<i>Streptocephalus rubicaudatus</i>	73700	1	n		2(3)	Crisinel et al 1994	Larver.
	<i>Streptocephalus sudanicus</i>	67750	2	n		2	Lahr et al 2001	Voksne
	<i>Streptocephalus texanus</i>	54600	1	n		RI2(3)	Crisinel et al 1994	Larver.
	<b><i>Thamnocephalus platyurus</i></b>	<b>25,05</b>	1	n			Ripley et al 2002	Larver
<b>Krebsdyr (Crustacea) Copepoda</b>	<i>Cyclops viridis</i>	1300	2	n			Konar et Ghosh 1981	
<b>Krebsdyr (Crustacea) Decapoda</b>	<b><i>Barytelphusa cunicularis</i></b>	<b>380</b>	4	n			Rao et Nagabhushanam 1987	
	<b><i>Macrobrachium lamarrei</i></b>	<b>130</b>	4	n	50		Nagabhushanam et al 1983	Intermolt
	<i>Macrobrachium lar</i>	851	2	n			Bajet et al 2012	
	<b><i>Macrobrachium rosenbergii</i></b>	<b>9</b>	4	n			Natarajan et al 1992	Post-larve
	<b><i>Orconectes nais</i></b>	<b>290</b>	1	n	95		Mayer et Ellersieck 1986	Larver

Overordnet systematiske gruppe	Art	EC <sub>50</sub> µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
	<i>Oziotelphusa senex ssp. senex</i>	6000	4	n	50		Victor 1989	Voksne
	<i>Palaemonetes kadiakensis</i>	12	4	n	95		Mayer et Ellersieck 1986	Voksne
	<i>Palaemonetes pugio</i>	9,06	4	n	"Technical grade"	2	Key et al 1998	Koncentrationerne var omkring 85% of nominelle
	<i>Palaemonetes pugio</i>	7,7	4	n			Key et Fulton 2006	Nyklækkede eller nyligt klækkede
	<i>Procambarus clarkii</i>	21000	4	y	96,8		FMC 2019: Brougher et al 2014, 1481 FYF	
Krebsdyr (Crustacea) Isopoda	<i>Asellus aquaticus</i>	32100	2	n	40		Frumin et 1992	Voksne
	<i>Caecidotea brevicauda</i>	6000	1	n	95		Mayer et Ellersieck 1986	Voksne
Krebsdyr (Crustacea) Ostracoda	<i>Cypridopsis vidua</i>	47	2	n	95		Mayer et Ellersieck 1986	Voksne
Insekter (Insecta) Hemiptera	<i>Anisops sardeus</i>	42,2	2	n		2	Lahr et al 2001	Voksne
	<i>Notonecta undulata</i>	110	2	n	94		Federle et Collins 1976	Voksne
Insekter (Insecta) Ephemeroptera	<i>Centroptilum triangulifer</i>	23	2	y	96,8		FMC 2019: Brougher et al 2014, 1442 FYF	
	<i>Cloeon sp.</i>	5,5	1	n			Cano et al 1999	Larver
	<i>Drunella grandis</i>	100	4	n			Gaufin et al 1965	Nymfer
	<i>Hexagenia sp.</i>	631	1	n	95		Carlson 1966	Naiad

Overordnet systematiske gruppe	Art	EC <sub>50</sub> µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
<b>Insekter (Insecta) Odonata</b>	<i>Lestes congener</i>	<b>27</b>	1	n	95		Mayer et Ellersieck 1986	Larver
	<i>Orthetrum albistylum ssp. speciosum</i>	730	2	n			Nishiuchi et al Asano 1978	
<b>Insekter (Insecta) Plecoptera</b>	<i>Claassenia sabulosa</i>	<b>13</b>	1	n	95		Mayer et Ellersieck 1986	
	<i>Hesperoperla pacifica = Acroneuria p.</i>	<b>7,7</b>	5	n	95	2	Jensen et Gaufin 1964a	niads, flow-through
	<i>Isoperla sp.</i>	<b>4,6</b>	1	n	95		Mayer et Ellersieck 1986	Naiad
	<i>Pteronarcella badia</i>	<b>10</b>	1	n	95		Mayer et Ellersieck 1986	
	<i>Pteronarcys californica</i>	<b>10</b>	4	n	95		Mayer et Ellersieck 1986	
<b>Insekter (Insecta) Trichoptera</b>	<i>Arctopsyche grandis</i>	<b>32</b>	4	n			Gaufin et al 1965	Larver
	<i>Hydropsyche californica</i>	<b>22,5</b>	4	n			Gaufin et al 1965	Larver
	<i>Hydropsyche sp.</i>	<b>12,3</b>	1	n	95		Carlson 1966	Larver
	<i>Limnephilus sp.</i>	<b>6,8</b>	1	n	95		Mayer et Ellersieck 1986	Larver
<b>Insekter (Insecta) Diptera</b>	<i>Aedes aegypti</i>	<b>5,3</b>	1	n	96	2	Failloux et al 1994	Larvere
	<i>Aedes albopictus</i>	<b>50</b>	1	n	99,2		Polnlawat et al 2005	Larver
	<i>Aedes atropalpus</i>	<b>172</b>	1	n	91		Cilek et al 1995	
	<i>Aedes cantans</i>	<b>48,8</b>	1	n			Rettich 1977	Larver

Overordnet systematiske gruppe	Art	EC <sub>50</sub> µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
	<i>Aedes communis</i>	38,2	1	n			Rettich 1977	Larver
	<i>Aedes excrucians</i>	30,3	1	n			Rettich 1977	Larver
	<i>Aedes hendersoni</i>	66	1	n	91		Cilek et al 1995	Larver
	<i>Aedes nigromaculis</i>	68	1	n			Mulla et al 1964	Larver
	<i>Aedes polynesiensis</i>	3,5	1	n	96	2	Failloux et al 1994	Larvere
	<i>Aedes punctor</i>	44,1	1	n			Rettich 1977	Larver
	<i>Aedes quadrimaculatus</i>	1	2	n		2	Milam et al 2000	Larvere
	<i>Aedes taeniorhynchus</i>	25,88	1	n			Montada et al 1994	Larver
	<i>Aedes triseriatus</i>	43	1	n	91		Cilek et al 1995	
	<i>Aedes trivittatus</i>	32,2	1	n	95		Rubio-Moran et al 1981	Larver
	<i>Aedes vexans</i>	26,1	1	n			Rettich 1977	Larver
	<i>Anopheles albimanus</i>	120	1	n	5		Kawada et al 1993	Larver
	<i>Anopheles coustani</i>	200		n			Hamed et al 1983	Larver
	<i>Anopheles gambiae</i>	190	1	n	5		Kawada et al 1993	Larver
	<i>Anopheles pseudopunctipennis</i>	2,27	1	n			Ocampo et al 2000	Larver

Overordnet systematiske gruppe	Art	EC <sub>50</sub> µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
	<i>Anopheles quadrimaculatus</i>	1	2	n		2	Milam et al 2000	
	<i>Anopheles stephensi</i>	4	1	n			Chitra et Pillai 1984	Larver
	<i>Atherix variegata</i>	1450	1	n	95		Mayer et Ellersieck 1986	Instar
	<i>Chironomus crassicaudatus</i>	56	1	n			Ali 1981	Larver
	<i>Chironomus decorus</i>	32	1	n			Ali 1981	Larver
	<i>Chironomus plumosus</i>	8,4	4	n			Vedamanikam 2009	Larver
	<i>Chironomus riparius</i>	0,44	1	n	97	2	Hoffman et Fisher 1994	Larver
	<i>Chironomus tentans</i>	3,5	2	y	96,8		FMC 2019: Brougher et al 2014, 1443 FYF	
	<i>Chironomus tepperi</i>	8,4	1	n			Stevens 1992	Larver
	<i>Chironomus utahensis</i>	3	1	n		2(3)	Ali et Mulla 1977	Instar .
	<i>Cricotopus sp.</i>	25	1	n		2(3)	Ali et Mulla 1977	Instar. Not reported as a formulation
	<i>Culex antennatus</i>	250		n			Hamed et al 1983	Larver
	<i>Culex fatigans</i>	16	1	n			Das et Rajagopalan 1979	Larver
	<i>Culex fuscocephala</i>	44,426	1	n			Vijayan et al 1993	Larver
	<i>Culex nigripalpus</i>	21,7	1	n			Boike et al 1989	Larver

Overordnet systematiske gruppe	Art	EC <sub>50</sub> µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
	<i>Culex pipiens</i>	1,5	1	n		2	Gaaboub et al 1975	4th instar
	<i>Culex pipiens</i>	1,6	2	n	95	2	Zahran et Abdelgaleil 2011	Fourth instar
	<i>Culex quinquefasciatus</i>	6,7	1	n	96	2	Failloux et al 1994	Larvere
	<i>Culex quinquefasciatus</i>	3,5	1	n	95	2	Aguilera et al 1995	Larver
	<i>Culex sitiens</i>	4,6	1	n			Komalamisra et al 2006	Larver
	<i>Culex tarsalis</i>	10	1	n			Ziegler et al 1987	Larver
	<i>Culex tritaeniorhynchus</i>	21	1	n			Shim& Self 1973	Larver
	<i>Culiseta annulata</i>	24,5	1	n			Rettich 1977	Larver
	<i>Culiseta longiareolata</i>	17	1	n			Cano et al 1999	Larver
	<i>Dicrotendipes californicus</i>	80	1	n			Ali et Mulla 1980	Larver
	<i>Glyptotendipes paripes</i>	4	1	n			Ali 1981	Larver
	<i>Goeldichironomus holoprasinus</i>	28	1	n			Ali 1981	Larver
	<i>Procladius sp.</i>	5	1	n		2(3)	Ali et Mulla 1977	Larver. I ECOTOX angivet at en formulering er brugt, men dette er

Overordnet systematiske gruppe	Art	EC <sub>50</sub> µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
								ikke angivet i artiklen
	<i>Simulium vittatum</i>	54,2	2	y	98		Overmyer et al 2003	Larver
	<i>Tanytarsus sp.</i>	32	1	n			Ali 1981	Larver
	<i>Toxorhynchites splendens</i>	49,8	2	n			Tietze et al 1993	Larver
Insekter (Insecta) Coleoptera	<i>Eretes sticticus</i>	430	2	n			Jeyasingam et al 1978	
	<i>Peltodytes sp.</i>	1500	2	n	94		Federle et Collins 1976	Voksne
Fish	<i>Alburnus alburnus</i>	3780	4	n	95		Guylas et Csanyi 1984	
	<i>Ameiurus melas</i>	18500	1	n	95		Mayer et Ellersieck 1986	
	<i>Anabas testudineus</i>	11800	4	n	95		Chandran et al 1991	
	<i>Anguilla japonica</i>	4100	2	n			Yokoyama et al 1988	
	<i>Aplocheilus lineatus</i>	975	2	n			Jacob et al 1982	
	<i>Barbonymus gonionotus</i>	6520	2	n	57		Hoque 2000	
	<i>Barbus barbus ssp. plebejus</i>	6600	1	n			Scirocchi et D'Erme 1980	
	<i>Barbus dorsalis</i>	3700	4	y	50		Rao et al 1967	
	<i>Barbus ticto</i>	13	2	n	96,45		Bhatia 1971	



Overordnet systematiske gruppe	Art	EC <sub>50</sub> µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
	<i>Barilius vagra</i>	7390	4	y	57		Nishiuchi et al Asano 1978	Voksne
	<i>Brachydanio rerio</i>	266	4	n	?		FMC 2019: Busquet et al 2014.	embryoner
	<i>Carassius auratus</i>	3150	4	y			Birge et al 1979	Æg
	<i>Channa orientalis</i>	16420	2	n	50		Verma et al 1984	
	<i>Channa punctata</i>	3890	4	n			Haider et Inbaraj 1986	Voksne
	<i>Cirrhinus mrigala</i>	880	4	n			Verma et al 1984	Larver
	<b><i>Clarias batrachus</i></b>	<b>12</b>	4	n			Sinha et al 1991	
	<i>Clarias gariepinus</i>	8220	4	n	57		FMC 2019:Ahmad 2012	
	<i>Colisa fasciata</i>	5360	2	n	50		Verma et al 1984	
	<i>Cyprinion watsoni</i>	7930	1	n	57		Khattak et Hafeez 1996	
	<b><i>Cyprinus carpio</i></b>	<b>85</b>	4	n			Verma et al 1981	Larver
	<b><i>Fundulus diaphanus</i></b>	<b>240</b>	4	n			Rehwoldt et al 1977	Unge fra samme år
	<i>Gambusia affinis</i>	2900	4	y	96,8	1 or 2	FMC 2019:Document 1426 FYF, 2014	
	<i>Gambusia holbrooki</i>	12780		n			Areekul,S. 1986	
	<i>Garra gotyla ssp. gotyla</i>	3500	4	n			Thakur et Sahai 1994	

Overordnet systematiske gruppe	Art	EC <sub>50</sub> µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
	<i>Gasterosteus aculeatus</i>	21,7	4	y	96	1 or 2	FMC 2019:Document 381 FYF, 2002	
	<i>Gasterosteus aculeatus</i>	46	4	y	96,8	1 or 2	FMC 2019:Document 881 FYF, 2011	
	<b><i>Gasterosteus aculeatus</i></b>	<b>31,6</b>					Geometrisk gennemsnit	
	<i>Gibelion catla</i>	2350	3	n	50		Arora 1971	
	<i>Gila elegans</i>	15300	4	y	93		Beyers et al 1994	Larver
	<b><i>Heteropneustes fossilis</i></b>	<b>11,798</b>	4	n	50		Dutta et al 1992	
	<i>Ictalurus furcatus</i>	17000	4	n	98,1		Aker et al 2008	"Fingerling" stadie
	<i>Ictalurus punctatus</i>	8900	2	y				
	<b><i>Jordanella floridae</i></b>	<b>0,235</b>	9	y	95		Hermanutz 1978	
	<i>Labeo fimbriatus</i>	8500	2	n	50		Sreenivasan Swaminathan 1967	
	<b><i>Labeo rohita</i></b>	<b>2</b>	4	n	50		FMC 2019: Mishra et al 2012	
	<i>Labeo rohita</i>	4,5	4	n	50		Patil et David 2010	"Fingerling" stadie
	<i>Lepidocephalichthys thermalis</i>	7750	2	n	50		Kumari et Nair 1978	Juvenile
	<b><i>Lepomis cyanellus</i></b>	<b>130</b>	4	y	96,8	1 or 2	FMC 2019:Document 1408 FYF, 2014	
	<b><i>Lepomis gibbosus</i></b>	<b>480</b>	4	n			Rehwoldt et al 1977	Unge fra samme år

Overordnet systematiske gruppe	Art	EC <sub>50</sub> µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
	<i>Lepomis macrochirus</i>	46	4	y	96,8	1 or 2	FMC 2019:Document 879 FYF, 2011	
	<i>Lepomis macrochirus</i>	54	4	y	96,9	1 or 2	FMC 2019:Document 314 FYF, 2001	
	<i>Lepomis macrochirus</i>	48	4	n	96,9		US EPA 1992	
	<b><i>Lepomis macrochirus</i></b>	<b>49</b>	4				Geometric mean of values above	
	<b><i>Lepomis microlophus</i></b>	<b>170</b>	1	n	95		Mayer et Ellersieck 1986	
	<i>Leucaspius delineatus</i>	7640	2	n			Ardo 1974	
	<i>Leuciscus cephalus</i>	1807	4	n	20		Verep 2006	
	<i>Melanotaenia fluviatilis</i>	2090	4	y			Reid et al 1995	Voksne
	<b><i>Micropterus salmoides</i></b>	<b>368</b>	1	n	95		Mayer et Ellersieck 1986	
	<i>Morone americana</i>	1067	4	n			Rehwoldt et al 1977	Unge fra samme år
	<b><i>Morone saxatilis</i></b>	<b>12</b>	4	y	94,2		Fujimura et al 1991	Larver
	<i>Mystus tengara</i>	8000	4	n	95		Chandran et al 1991	
	<i>Mystus vittatus</i>	2000	4	n	50		Verma et al 1984	
	<i>Nemacheilus angorae</i>	3023,7	4	n			Gul et al 2007	Voksne
	<i>Nothobranchius guentheri</i>	2900	1	y			Shedd et al 1999	Små unger

Overordnet systematiske gruppe	Art	EC <sub>50</sub> µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
	<b><i>Notopterus notopterus</i></b>	<b>77</b>	4	n			Gupta et al 1994	
	<b><i>Oncorhynchus clarkii</i></b>	<b>150</b>	4	n	95		Post et Schroeder 1971	
	<i>Oncorhynchus kisutch</i>	720	4	y	96,8	1 or 2	FMC 2019:Document 1445 FYF, 2014	
	<i>Oncorhynchus kisutch</i>	226	1	n	95		Mayer et Ellersieck 1986	
	<b><i>Oncorhynchus kisutch</i></b>	<b>403</b>					Geometrisk gennemsnit	
	<i>Oncorhynchus mykiss</i>	39	1	n	95		Mayer et Ellersieck 1986	
	<i>Oncorhynchus mykiss</i>	180	4	y	96,9	1 or 2	FMC 2019:Document 306 FYF, 2001	
	<b><i>Oncorhynchus mykiss</i></b>	<b>84</b>					Geometrisk gennemsnit	
	<i>Oncorhynchus tshawytscha</i>	364	4	y	98,7		FMC 2019:Detrich et al 2014. 11°C	Udført ved 11°C. Foretrukne temperatur 0,7-8°C
	<i>Oreochromis mossambicus</i>	358,9	2	n	50		Sailatha et al 1981	
	<i>Oreochromis niloticus</i>	140	4	n	80		Liong et al 1988	
	<i>Oryzias latipes</i>	1500	4	y	96,8	1 or 2	FMC 2019:Document 1407 FYF, 2014	
	<i>Oryzias latipes</i>	1800	2	n	95		Tsuda et al 1997	
	<i>Pangasianodon hypophthalmus</i>	7,76	4	n	50		FMC 2019: Shashikumar et al 2018	Ved 30°C. Ved 32° and 34° var LC50 henholdsvis 6,5 og 4,5 µg/l.
	<i>Pelophylax ridibundus</i>	29000	4	n	25		Sayim 2008	Haletudser

Overordnet systematiske gruppe	Art	EC <sub>50</sub> µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
	<i>Perca flavescens</i>	482	1	n	95		Mayer et Ellersieck 1986	
	<i>Pimephales promelas</i>	28000	4	y	96,8	1 or 2	FMC 2019:Document 1380 FYF, 2013	
	<i>Pimephales promelas</i>	27000	4	y	99,1	1 or 2	FMC 2019:Document 1381 FYF, 2013	
	<i>Pimephales promelas</i>	12400	1	n	95		Mayer et Ellersieck 1986	
	<i>Pimephales promelas</i>	21085					Geometric mean of values above	
	<i>Poecilia reticulata</i>	880	4	n	100		Pickering et al 1962	
	<i>Pseudorasbora parva</i>	14500	4	n	98		Li et Fan 1996	
	<i>Pseudosphromenus cupanus</i>	4594	2	n			Jacob et al 1982	
	<i>Ptychocheilus lucius</i>	9140	4	y	93		Beyers et al 1994	Larver
	<i>Puntius sophore</i>	1650	4	n	30		Khangarot et Ray 1988	
	<i>Puntius stigma</i>	9560	2	n			Verma et al 1984	
	<i>Rasbora daniconius</i>	4000	4	n	95		Singh et al Sahai 1984	
	<i>Rhodeus sericeus ssp. amarus</i>	5060	4	n	95		Guylas et Csanyi 1984	
	<i>Rutilus rutilus</i>	12000	4	n			Scirocchi et D'Erme 1980	
	<i>Salmo salar</i>	33	2	n			Keenleyside 1958	Unge

Overordnet systematiske gruppe	Art	EC <sub>50</sub> µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
	<i>Salmo trutta</i>	128	1	n	95		Mayer et Ellersieck 1986	
	<i>Salvelinus fontinalis</i>	120	4	n	95		Post et Schroeder 1971	
	<i>Salvelinus namaycush</i>	99	1	n	95		Mayer et Ellersieck 1986	
	<i>Sander vitreus</i>	11	1	n	95		Mayer et Ellersieck 1986	
	<i>Tilapia sp.</i>	45,99	4	n			Bajet et al 2012	"Fingerling" stadie
	<i>Tinca tinca</i>	16200	3	n			Scirocchi et D'Erme 1980	
	<i>Trichogaster pectoralis</i>	983,2	4	n	57		Mohsin et al 1984	
	<i>Umbra pygmaea</i>	240	4	y	99,5	2	Bender et Westman 1976	
	<i>Zacco platypus</i>	9700	1	n			Shim &Self 1973	
<b>Amphibia</b>	<i>Bufo americanus</i>	7968	4	y	50,6	2	Relyea 2004	Haletudser. Værdien skønnet ud fra fig. 1
	<i>Bufo woodhousei ssp. fowleri</i>	1900	1	n	95		Mayer et Ellersieck 1986	Haletudser
	<i>Euphlyctis cyanophlyctis</i>	3523	4	n	50		Giri et 2012	Haletudser
	<i>Hoplobatrachus tigerinus</i>	170	6	n			Abbasi et Soni 1991	Haletudser
	<i>Hyla versicolor</i>	8751	4	y	50,6	2	Relyea 2004	Haletudser, Værdien skønnet ud fra fig. 1
	<i>Lithobates palustris = Rana p.</i>	16400	2	y	"Pure"	2	Budischak et al 2009	Haletudser. Geometrisk gennemsnit af nederste og øverste værdi i

Overordnet systematiske gruppe	Art	EC <sub>50</sub> µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
								spandet givet i artiklen
	<i>Pseudacris triseriata</i>	560	1	n	95		Mayer et Ellersieck 1986	Haletudser
	<i>Rana boylii</i>	2137	4	n	99		Sparling et Fellers 2007	Haletudser
	<i>Rana catesbeiana</i>	5357	4	y	50,6	2	Relyea 2004	Haletudser, Værdien skønnet ud fra fig. 1
	<i>Rana clamitans = Lithobates c.</i>	5323	4	y	50,6	2	Relyea 2004	Haletudser, Værdien skønnet ud fra fig. 1
	<i>Rana hexadactyla = Euphlyctis h.</i>	0,59	4	n	50	2	Khangarot et al 1985	Haletudser
	<i>Rana limnocharis</i>	2271	2	n	50		Pan,D.Y. et Liang 1993	Haletudser
	<i>Rana pipiens = Lithobates p.</i>	4842	4	y	50,6	2	Relyea 2004	Haletudser, Værdien skønnet ud fra fig. 1
	<i>Rana sylvaticus = Lithobates s,</i>	5204	4	y	50,6	2	Relyea 2004	Haletudser, Værdien skønnet ud fra fig. 1
	<i>Rhinella arenarum</i>	19200		n			Venturino et al 1992	Larver
	<i>Xenopus laevis</i>	790	4	n	90		Snawder et Chambers 1989	Embryo

**Ferskvandsorganismer**  
 Kronisk giftighed  
*Freshwater, chronic toxicity*

Overordnede systematiske grupper	Art	EC10 µg/l	NOEC µg/l	Valgte værdi	Varighed dage	Målt (Yes/no)	Effekt	Renhed %	RI	Reference	Kommentarer
<b>Cyano-bacteria</b>	<i>Anabaena flos-aquae</i>	27900		<b>27900</b>	3	y	Vækstrate	96	RI2	FMC 2019: Dobbins, Kendall, Porch 2012	
<b>Alger (Algae) Chlorophyta</b>	<i>Raphidocelis subcapitata</i>	4000			3	y	Vækstrate	96,4	RI2	FMC 2019: Jenkins 1993	
	<i>Raphidocelis subcapitata</i>	6500			3	y	Vækstrate	96	RI2	FMC 2019: Dobbins, Kendall and Porch 2012;	
	<i>Raphidocelis subcapitata</i>		1200		2	y	Tæthed af celler	99	RI2	Yeh & Chen 2006	
	<i>Raphidocelis subcapitata</i>	8500			2	n	Vækstrate	95		Kusk et al 2018	
	<i>Raphidocelis subcapitata</i>			<b>4035</b>						Geometrisk gennemsnit af værdierne ovenfor	
<b>Alger (Algae) Bacillariophyta</b>	<i>Navicula pelliculosa</i>	15400		<b>15400</b>	3	y	Growth rate	96	RI2	FMC 2019: Dobbins, Kendall, Porch 2012	
<b>Macro-phyta</b>	<i>Lemna gibba</i>	19900		<b>19900</b>	7	y	Growth rate	96	RI2	FMC 2019: Dobbins, Kendall, Porch 2012	
<b>Fladorme (Platyhelminthes) Trematoda</b>	<i>Echinoparyphium sp.</i>	7,9		<b>7,9</b>	6 h	y, initial	cercaria survival	"commercial grade"	RI2	Hua et al. 2016	EC10 blev estimeret ud fra figur 1. Eksponering i 6 timer er rimelig da cercarierne ikke ovelever i meget mere end 8 timer, hvis de ikke finder en vært.



Overordnede systematiske grupper	Art	EC10 µg/l	NOEC µg/l	Valgte værdi	Varighed dage	Målt (Yes/no)	Effekt	Renhed %	RI	Reference	Kommentarer
<b>Nema-tomorpha</b>	<i>Chordodes nobilii</i>	0,15		<b>0,15</b>	2	Initial	infection effectivitet	####	RI2	Achiorno et al 2009	De præparasitiske larver blev eksponeret for malathion i 48 timer og derefter sat sammen med Aedes aegyptii i 72 timer og infektionen observeret. Testmedier blev delvist udskiftet en gang i døgnet.
<b>Krebsdyr (Crustacea)</b>  <b>Branchiopoda</b>	<i>Ceriodaphnia dubia</i>	8*10 <sup>-04</sup>	0,001	<b>8E-04</b>	7	y	number of offsprings	1103 g/L	2	FMC 2019: V.C. Franco Perina 1995	EC <sub>10</sub> blev beregnet ved ekstrapolering mellem antallet af unger ved 0,00056 µg/l. Antallet af unger ved 0,00056 µg/l blev sat lig det geometriske gennemsnit af antallet i kontrollen, antallet ved 0,00032 µg/l og antallet ved 0,00056 µg/l, fordi det først er efter 0,00056 µg/l, at der sker et fald i antallet af unger.
	<i>Daphnia magna</i>	0,079	0,06	0,079	21	y	juveniles/fe male reproductive days	94	2	FMC 2019: G. Blakemore, D. Burgess 1990	Forfatterne beregnede en EC10 på 0,085 µg/l. Men Analyse af sammenhængen

Overordnede systematiske grupper	Art	EC10 µg/l	NOEC µg/l	Valgte værdi	Varighed dage	Målt (Yes/no)	Effekt	Renhed %	RI	Reference	Kommentarer
											resulterede i en EC10 = 0,079 µg/l. Spearman rank korrelationen var rs = -1,000 and product moment korrelation var r = -0,994, med P-værdier på henholdsvis [N = 6] 0,01 and 0,002 (tosidet)
	<i>Daphnia magna</i>		0,57	0,57	21	y	Young produced	99,8	2	Ding et al 2019	EC20 var 1,13 µg/l. Dette er ≈ LOEC. NOEC blev beregnet som 1,13/2.
	<i>Daphnia magna</i>	0,0047		0,0047	21	n	Young produced	50	2	Toumi et al 2015	
	<i>Daphnia magna</i>			<b>0,06</b>						Geometric mean of values above	
<b>Krebsdyr (Crustacea)</b> <b>Decapoda</b>	<i>Neocaridina denticulata</i>		7,2	<b>7,2</b>	28	y	survival	99,8	2	Ding et al 2019	EC <sub>20</sub> var 14,3 µg/l. Dette ≈ LOEC. NOEC blev beregnet ved 14,3/2.
<b>Insekter (Insecta)</b> <b>Plecoptera</b>	<i>Acroneturia pacifica = Hesperoperla p.</i>	0,54		<b>0,54</b>	30	n	naiads	95	2	Jensen & Gaufin 1964b	Beregnet fra data givet i figuren
	<i>Pteronarcys californica</i>	3,5		<b>3,5</b>	30	n	naiads	95	2	Jensen & Gaufin 1964b	Beregnet fra data givet i figuren
<b>Fisk (Fish)</b>	<i>Brachydanio rerio</i>	1994		<b>1994</b>	5	y	Hatching	99	2	Cook et al. 2005	Koncentrationerne blev målt hver anden dag. EC10 for overlevelse var omkring 2000 µg/l. EC10 for øjediameter var lavere, men det er svært at relatere dette til effekt på populationer

Overordnede systematiske grupper	Art	EC10 µg/l	NOEC µg/l	Valgte værdi	Varighed dage	Målt (Yes/no)	Effekt	Renhed %	RI	Reference	Kommentarer
	<i>Gila elegans</i>		990	<b>990</b>	32	y	weight of Larvere		RI2	Beyers et al. 1994	Forfatterne antager at der findes en tærskel værdi. Der er ikke givet nok data til kontrollere beregningerne og graden af effekt ved NOEC kan ikke vurderes. Koncentrationerne er blevet målt en gang om ugen, men det er ikke angivet om beregningerne er baseret på startkoncentrationen eller hvad.
	<i>Oncorhynchus mykiss</i>	9,2	21	<b>9,2</b>	97 days, 60 days posthatch	y	survival	94	2	FMC 2019: "1989; 034 FYF"	FMC 2019 udtaler, at "goodness of fit" er lav, og at man derfor har valgt NOEC i stedet for LC10. Men korrelationen mellem koncentration og overlevelse er statistisk signifikant ( $r_s = -0,943$ , $P = 0,02$ , tosidet og $r = 0,982$ , $P < 0,001$ , tosidert; $N = 6$ ). Endvidere er

Overordnede systematiske grupper	Art	EC10 µg/l	NOEC µg/l	Valgte værdi	Varighed dage	Målt (Yes/no)	Effekt	Renhed %	RI	Reference	Kommentarer
											usikkerhederne omkring NOEC snarere større, man har bare ikke mulighed for at se det.
	<i>Pimephales promelas</i>		80	<b>80</b>	21	y	Weight	96	2	EFSA, 2019: "2011; 1058 FYF"	Svarer til ca. 9% effekt
	<i>Pseudorasbora parva</i>		237	<b>237</b>	28	y	survival	99,8	2	Ding et al 2019	EC20 var 473 µg/l. This ≈ LOEC. NOEC NOEC blev beregnet ved 473/2
	<i>Ptychocheilus lucius</i>		1680	<b>1680</b>	32	y	survival and weight of Larvere		RI2	Beyers et al. 1994	Forfatterne antager at der findes en tærskel værdi. Der er ikke givet nok data til kontrollere beregningerne og graden af effekt ved NOEC kan ikke vurderes. Koncentrationerne er blevet målt en gang om ugen, men det er ikke angivet om beregningerne er baseret på startkoncentrationen eller hvad.
Padder (Amphibia)	<i>Rana catesbeiana</i>	263		<b>263</b>	28	n	Tadpoles, days to first	96	RI2	Fordham et al 2001	Koncentrationerne i stamopløsningerne blev målt, og

Overordnede systematiske grupper	Art	EC10 µg/l	NOEC µg/l	Valgte værdi	Varighed dage	Målt (Yes/no)	Effekt	Renhed %	RI	Reference	Kommentarer
							recorded loss of equilibrium				testmedierne blev udskiftet dagligt.
	<i>Xenopus laevis</i>		59	<b>59</b>	14	y	Wet weight	96	RI2	FMC 2019: "2011; 1059 FYF"	Svarer til ca. 9,2% effekt
	<i>Xenopus laevis</i>	not applicable	not applicable		21					FMC 2019: "2011; 1060 FYF"	Der blev ikke set en dosis-respons
	<i>Xenopus laevis</i>	1294			7	n	embryonal malformation	99	RI2	Bonfanti et al. 2004	Testmediet blev fornyet dagligt

**Saltvandsorganismer**  
 Akut giftighed  
*Saltwater, acute toxicity*

Overordnet systematisk gruppe	Art	EC50 µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
<b>Algae</b>	<i>Skeletonema costatum</i>	15110	3	y	96		FMC 2019: Dobbins et al 2012, 1263 FYF,	
<b>Rotifera</b>	<i>Brachionus plicatilis</i>	67000	1	y	95		Guzzella et al 1997	Neonate
<b>Bløddyr (Mollusca)</b>							FMC 2019: Brougher 2014, 1418 FYF	
<b>Bivalvia</b>	<i>Crassostrea virginica</i>	2700	4	y	96,8			
	<i>Kataysia opima</i>	6	4	n	50		Akarte et al 1986	
	<i>Mytilus edulis</i>	13400	2	y	95		Liu et Lee 1975	Æg
	<i>Mytilus galloprovincialis</i>	6000	7	n			Rao et Mane 1978	
<b>Krebsdyr (Crustacea)</b>							Leight,A.K., and R.F. Van Dolah 1999	
<b>Amphipoda</b>	<i>Gammarus palustris</i>	5,85	3	n				
<b>Krebsdyr (Crustacea)</b>							Wilkins et Metcalfe 1993	Larver
<b>Anostraca</b>	<i>Artemia salina</i>	37030	1	n				
<b>Krebsdyr (Crustacea)</b>							FMC 2019: Brougher et al 2014, 1427 FYF	
<b>Mysida</b>	<i>Americamysis bahia</i>	4,8	4	y	96,8			
	<i>Americamysis bahia</i>	2,2	4	y	94		FMC 2019: Forbis 1990, 036 FYF	
	<i>Americamysis bahia</i>	2,1	4	y	94		FMC 2019: Burgess 1989, 19 FYF	

Overordnet systematisk gruppe	Art	EC50 µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
	<i>Americamysis bahia</i>	1,96	4	n	94		US EPA 1992	
	<i>Americamysis bahia</i>	3	4	y	71,5	2	Goodman et al 1988	
	<b><i>Americamysis bahia</i></b>	<b>2,6</b>					Geometric mean of values above	
	<b><i>Neomysis mercedis</i></b>	<b>1,4</b>	4	y	94,2	2	Brandt et al. 1993	neonate, geometrisk gennemsnit af 2,2; 1,5; 1,4
	<b><i>Neomysis mercedis</i></b>	<b>1,4</b>	4	y	94,2		Brandt et al 1993	Neonate
Krebsdyr (Crustacea) Copepoda	<b><i>Tigriopus brevicornis</i></b>	<b>7,2</b>	4	n			Forget et al 1998	Nauplii
Krebsdyr (Crustacea) Decapoda	<b><i>Cancer magister</i></b>	<b>1,2</b>	4	n	95	2	Caldwell 1977	Zoea Larver. Testmedier fornyet dagligt
	<b><i>Crangon septemspinosa</i></b>	<b>33</b>	4	n			Eisler 1969	
	<b><i>Farfantepenaeus duorarum</i></b>	<b>12,5</b>	2	y			Bahner et Nimmo 1975	Unge voksne
	<b><i>Homarus americanus</i></b>	<b>3,7</b>	2	y	98	2	Zulkosky et al 2005	Larver
	<i>Litopenaeus stylirostris</i>	34197	2	n			Reyes et al 2002	Larver
	<b><i>Litopenaeus vannamei</i></b>	<b>21,46</b>	4	n		2	Salame et al 1996	Post-Larve
	<i>Metapenaeus monoceros</i>	1120	4	nnn	95		Reddy et Rao 1992	Intermolt
	<b><i>Pagurus longicarpus</i></b>	<b>83</b>	4	n	95		Eisler 1969 Reddy et Rao 1992 Salame et al 1996	Intermolt

Overordnet systematisk gruppe	Art	EC50 µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
	<b><i>Palaemon macrodactylus</i></b>	<b>9</b>	4	n	95 95		Schoettger 1970	Intermolt
	<i>Palaemonetes pugio</i>	7,7	4	n	95	2	Key et al 19981969	Nyklækkede
	<i>Palaemonetes vulgaris</i>	<b>82</b>	4	n	95	2	Eisler 1969Key et al 1998Schoettger 1970	Nyklækkede
	<i>Palaemonetes pugio</i>	9,1	4	y	"Technical grade"	2 2	Key 1995	Nyklækkede
	<b><i>Palaemonetes pugio</i></b>	<b>8,4</b>					Geometrisk gennemsnit	
	<b><i>Penaeus monodon</i></b>	<b>392</b>	4	n		2	Rao et al 1988	Intermolt
	<b><i>Uca marionis</i></b>	<b>60</b>	1	n		2	Yeragi,S.G., and V.A. Koli	
<b>Insekter (Insecta) Diptera</b>	<i>Culicoides sp.</i>	87,9	1	n			Kline et al 1985	Larver
<b>Pighude (Echinodermata)</b>	<i>Echinometra mathaei</i>	9790	4	n			Verma et al 1982	
<b>Fisk (Fish)</b>	<b><i>Anguilla rostrata</i></b>	<b>829</b>	4	n			Eisler 1970Verma et al 1982Kline et al 1985	
<b>Fisk (Fish)</b>	<b><i>Aphanius fasciatus</i></b>	<b>348</b>	2	n			Boumaiza et al 1979	
<b>Fisk (Fish)</b>	<b><i>Barbus stigma</i></b>	<b>19,5</b>	4	n			Khillare et al Wagh 1988	
	<i>Cyprinodon variegatus</i>	47,4	4	y	96,8	1 or 2	FMC 2019:Document 1039 FYF, 2011	
	<i>Cyprinodon variegatus</i>	1354	4	y	96,8	1 or 21	FMC 2019:Document 880 FYF,	
	<i>Cyprinodon variegatus</i>	51	4	y		2	Hansen & Parrish 1977	Juvenile



Overordnet systematisk gruppe	Art	EC50 µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
	<i>Cyprinodon variegatus</i>	40	4	y	94	1 or 2	FMC 2019:Document 018 FYF, 1989	
	<b><i>Cyprinodon variegatus</i></b>	<b>60</b>						Geometrisk gennemsnit af værdierne ovenfor Juvenile
	<b><i>Fundulus heteroclitus</i></b>	<b>22,51</b>	4	ny	50	1 or 2	Scott et al 1987	
	<b><i>Fundulus majalis</i></b>	<b>250</b>	4	nn			Eisler 1970	
	<b><i>Fundulus similis</i></b>	<b>150</b>	2	n	95		Hansen 1980	
	<i>Gasterosteus aculeatus</i>	76,9	4	n	57		Katz 1961	
	<i>Leiostomus xanthurus</i>	550	2	n			Butler 1964	Juvenile
	<b><i>Menidia beryllina</i></b>	<b>30</b>	4	n			Lewis 1993	Juvenile
	<b><i>Menidia menidia</i></b>	<b>125</b>	4	n			Eisler 1970	Juvenile
	<b><i>Mugil cephalus</i></b>	<b>330</b>	2	n	95		Hansen 1980	
	<b><i>Mugil curema</i></b>	<b>570</b>	2	n			Butler 1963	Juvenile
	<b><i>Oncorhynchus tshawytscha</i></b>	<b>33,7</b>	4	n	95		Schoettger 1970	
	<i>Rivulus marmoratus</i>	1100	4	n			Lewis 1993	
	<i>Sphoeroides maculatus</i>	3250	4	n			Eisler 1970	

Overordnet systematisk gruppe	Art	EC50 µg/l	Varighed dage	Målt yes/no	Renhed %	RI	Reference	Kommentarer
	<i>Thalassoma bifasciatum</i>	27	4	n			Eisler 1970	

## Saltvandsorganismer

Kronisk giftighed

*Saltwater, chronic toxicity*

Overordnede systematiske grupper	Art	EC10 µg/l	NOEC µg/l	Valgte værdi	Varighed dage	Målt (Yes/no)	Effekt	Renhed %	RI	Reference	Kommentarer
<b>Alger (Algae) Bacillariophyceae</b>	<i>Skeletonema costatum</i>	5690		<b>5690</b>	3	y	Growth rate	96	2	FMC 2019: Dobbins, Kendall, Porch 2012	
<b>Krebsdyr (Crustacea) Mysida</b>	<i>Americamysis bahia</i>	0,295		<b>0,295</b>	39	y	juveniles/day	96	RI2	FMC 2019: M.B. Claude, T.Z. Kendall, S.P. Gallagher, H.O. Krueger 2012	
<b>Krebsdyr (Crustacea) Decapoda</b>	<i>Cancer magister</i>		0,2	<b>0,2</b>	69	n	zoea survival	95	2	Caldwell 1977	Koncentrationerne blev ikke målt, men testmedier blev udskiftet tre gange om ugen. Der ser ud til at være en NOEC for forsinkelse af hamskifte i sidste zoea stadie på 0,02 µg/l, men det er ikke overbevisende
<b>Fisk (Fish)</b>	<i>Cyprinodon variegatus</i>	7,9	8,2	<b>7,9</b>	28 days post hatch	y	survival		2	FMC 2019: "2011; 1072 FYF"	FMC 2019 udtaler, at "goodness of fit" er lav, og at man derfor har valgt NOEC i stedet for LC10. Men korrelationen mellem koncentration og overlevelse er statistisk signifikant ( $r_s = -0,883$ , $0,02 < P < 0,05$ , two-tailed and

Overordnede systematiske grupper	Art	EC10 µg/l	NOEC µg/l	Valgte værdi	Varighed dage	Målt (Yes/no)	Effekt	Renhed %	RI	Reference	Kommentarer
											r = -0,965, P<0,001 ). Endvidere er usikkerhederne omkring NOEC snarere større, man har bare ikke mulighed for at se det.
	<i>Cyprinodon variegatus</i>				10 (?) weeks	y	survival		2 (==>4)	Hansen & Parrish 1977	Oplysningerne om udførelsen af testene er mangelfulde, så det kan være lidt svært at vurdere forsøget (tenderer mod RI4). Da studiet ovenfor er betydeligt mere troværdigt bruges det.

## Fugle og pattedyr

### Birds and mammals

Det antages at både kaninerne, rotterne og fuglene i forsøgene er blevet fodret med kommercielt foder ("commercial fodder" i tabel 8 i vejledningen), da det er nemmere at få blandet stoffet i et sådant standardfoder. I rapporterne nævnes også standardfoder og "basal ration".

*It is assumed that both the mammals and the birds in the studies have been fed "commercial fodder". This because it is more convenient to mix the substance into such a standard diet. The reports mention standard diets and "basal ration".*

Overordnet systematiske grupper	Art	Energi normaliseret EC10 eller NOEC, mg/kJ	Valgte værdi	EC10, mg/kg foder	NOEC, mg/kg foder	NOAEL, mg/kg lgv per dag	Dage	Renhed, %	Reference	Kommentarer
<b>Fugle (Aves)</b>	Colinus virginianus	0,0079	110		110	13,5	147	96,4	FMC 2019: 103 FYF 1995	21 uger. Regressing ovaries etc.
	Anas platyrhynchos	0,086	1200		1200		32	94	FMC 2019: 069 FYF 1993	Reproduktion
	Colinus virginianus	0,022	300		300		147	94	FMC 2019: 029 FYF	Reproduktion

Overordnet systematiske grupper	Art	Energi normaliseret EC10 eller NOEC, mg/kJ	Valgte værdi	EC10, mg/kg foder	NOEC, mg/kg foder	NOAEL, mg/kg lgv per dag	Dage	Renhed, %	Reference	Kommentarer
	<i>Colinus virginianus</i>	0,016	218	218			28	94	FMC 2019: 038 FYF 1989	Overlevelse. FMC 2019 hævder at "Compliance with the OECD 206 validity criteria could not be verified" og at studiet derfor var ikke troværdigt. Men kontroldødelighed var mindre end 10% (1 fugl) og udover det var der ingen tegn på effekter i kontrollen. Der var endvidere en statistisk signifikant dosis-respons. LC10 blev beregnet ud fra data givet i rapporten
<b>Aves</b>	<i>Phasianus colchinus</i>	0,0018	25,14		25,14	2514 (LC50)				
	Species	Energy normalized EC10 or NOEC, mg/kJ	Chosen value	EC10, mg/kg feed	NOEC, mg/kg feed	NOAEL, mg/kg bw per day	Days	Purity, %	Reference	Comments
<b>Mammalia</b>	<i>Oryctolagus cuniculus</i>	0,060	832,5		832,5	25	29	92,4	FMC 2019: 008 FYF 1985	Reproduction. NOEC calculated from NOAEL with factor 33,3

Overordnet systematiske grupper	Art	Energi normaliseret EC10 eller NOEC, mg/kJ	Valgte værdi	EC10, mg/kg foder	NOEC, mg/kg foder	NOAEL, mg/kg lgv per dag	Dage	Renhed, %	Reference	Kommentarer
	<i>Oryctolagus cuniculus</i>	0,12	1665		1665	50	29	96	FMC 2019: TOX-FYF 4597 2018	
	<i>Rattus norvegicus</i>	0,0014	20		20	1			FMC 2019: 165 FYF 1996	Reproduction. NOEC calculated from NOAEL with factor 20
	<i>Rattus norvegicus</i>	0,0029	40			EL10: 400	20	96	FMC 2019: TOX-FYF4595 2018	EC10 beregnet ud fra data i rapporten og med en usikkerhedsfaktor på 10 for at tage højde for, at det er et korttidsforsøg
	<i>Rattus norvegicus</i>	0,0095	132		132	1700	20	96	FMC 2019: TOX-FYF041 2018	NOAEL og NOEC givet i rapporten. Standard ville være at dividere NOAEL med 10 for at skønne NOEC

## Bilag B

NOEC værdier i muslinger.

Muslinger er valgt som kritiske fødekilde, da malathion ikke opkoncentreres gennem fødekæden. Jævnfør tabel 7 i EU vejledningen (EU 2018a) antages energiindholdet i muslinger at være 19 kJ/g<sub>tørvægt</sub> og vandindholdet 92% (0,92). Værdierne normaliseres ikke til lipidindholdet. Valgte værdier er mærket med fed skrift.

*NOEC values in mussels.*

*Mussels are chosen as critical diet because malathion does not biomagnify. The energy content of mussels is set at 19 kJ/g<sub>dw</sub> and the moisture content at 92%. Normalization to lipid content has not been employed as the lipid content of mussels is low and the substance isn't especially hydrophobic (water solubility is 143 mg/l).*

NOEC (mg/kg tørvægt) i muslinger =  
 energinormaliseret NOEC<sub>lab-foder</sub>\*energi i muslinger\*(1-vandindhold i muslinger)

Art/species	NOEC i muslinger/mussels mg/kg <sub>dw</sub>		Studie i FMC 2019
<i>Colinus virginianus</i>	12		103 FYF
<i>Colinus virginianus</i>	33		029 FYF
<i>Colinus virginianus</i>	24		038 FYF
Geometrisk gennemsnit	<b>21</b>		
<i>Anas platyrhynchos</i>	<b>131</b>		069 FYF
<i>Phasianus colchinus</i>	<b>2,7</b>		1249 FYF
<i>Oryctolagus cuniculus</i>	91		008 FYF
<i>Oryctolagus cuniculus</i>	182		4597 FYF
<i>Rattus norvegicus</i>	2,2		165 FYF
<i>Rattus norvegicus</i>	44		041 FYF
<i>Rattus norvegicus</i>	14		4595 FYF
Geometrisk gennemsnit	<b>11</b>		



# Bilag C

Udtræk for malathion fra den danske QSAR database ([Danish \(Q\)SAR Database \(dtu.dk\)](http://qsar.food.dtu.dk)).

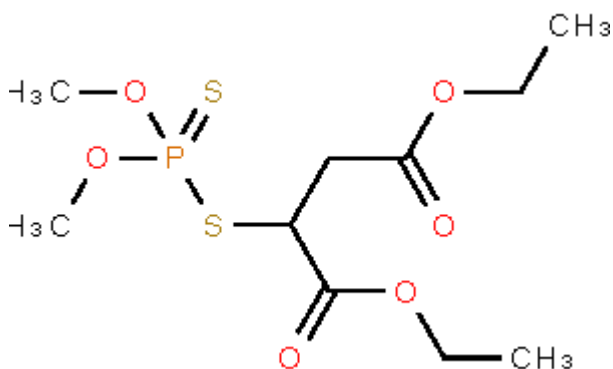
Databasen udvikles og vedligeholdes af Danmarks Tekniske Universitet.

Danish (Q)SAR Database, <http://qsar.food.dtu.dk>

Date: 09-11-2022

## (Q)SAR predicted profile

Structure (as used for QSAR prediction):



SMILES (used for QSAR prediction): C(=O)C(CC(=O)OCC)SP(=S)(OC)OC

ID

REACH EC Number (pre-registration, by 2013)	204-497-7	REACH EC Number (registration, by Dec. 2019)	
Registry Number	121-75-5	PubChem CID	
EU CLP Harmonized Classification*	Acute Tox. 4 *; Skin Sens. 1; Aquatic Acute 1; Aquatic Chronic 1	DK-EPA / DTU QSAR-based CLP Advisory Classification	
REACH registration cumulated minimum annual tonnage		US TSCA (Oct. 2021)	
Tox21 (2019)	Yes	ToxCast (Oct. 2021)	Yes
Molecular Formula	C <sub>10</sub> H <sub>19</sub> O <sub>6</sub> P <sub>1</sub> S <sub>2</sub>	Molecular weight (g/mole)	330.35
Chemical Name	malathion		

(Annex VI to CLP up to and including the 9th ATP, and including Nordic Council of Minister SPIN list for group entries)

### Melting point, Boiling point and Vapour pressure

Melting Point (deg C)	-23.58	Melting Point Experimental (deg C)	
Boiling Point (deg C)	351.17	Boiling Point Experimental (deg C)	156-157 @ 0.7 mm Hg
Vapour Pressure (atm)	EPI.Estimated_VP_atm	Vapour Pressure Experimental (atm)	EPI.Exp_VP_atm
Vapour Pressure (mm Hg)	0.000124	Vapour Pressure Experimental (mm Hg)	3.38E-06
Vapour Pressure (Pa)	0.01653	Vapour pressure Subcooled Liquid (Pa)	

*EPI MPBPVP models*

### Henry's Law Constant

HLC Bond Method (atm-m <sup>3</sup> /mole)	8.392E-010	HLC Group Method (atm-m <sup>3</sup> /mole)	
HLC Via VP/WSol (atm-m <sup>3</sup> /mole)	6.871E-007	HLC Via VP/WSol (Pa-m <sup>3</sup> /mole)	0.06962
Henry's Law Const. Exp db (Pa-m <sup>3</sup> /mole)	0.000495	Henry's Law Const. Exp db (atm-	4.89E-009

*EPI HENRYWIN models*

### Water Solubility

Water solubility from Kow (mg/L)	78.45	Water solubility from Fragments (mg/L)	427.53
Water solubility Exp (mg/L)	143	Water solubility Exp Ref	BOWMAN,BT & SANS,WW (1983)

*EPI WATERNT model*

### Hydrolysis

Hydrolysis Ka half-life pH 7		Hydrolysis Kb half-life pH 7	1324.762
Hydrolysis Ka half-life pH 8		Hydrolysis Kb half-life pH 8	132.459

*EPI HYDROWIN model*

### pKa

pKa Acid	999
- Standard deviation (±)	0
pKa Base	-999
- Standard deviation (±)	0

*ACDLabs model*

*pKa estimate 999: no acidic moiety found. pKa estimate -999: no basic moiety found.*

Partition coefficients

pH	1	4	5	6	7	8	9
LogD	2.41	2.41	2.41	2.41	2.41	2.41	2.41

Minimum LogD in the pH interval 4-9	2.41	Maximum LogD in the pH interval 4-9	2.41
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*ACDLabs models*

*LogD: Log octanol-water partition coefficient, which for ionizable compounds varies with the pH-dependent amounts of neutral and ionized species*

Log Koa	9.059	Log Kaw	-6.699
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*EPI KOAWIN models*

*Koa: octanol-air partition coefficient. Kaw: air-water partition coefficient.*

Log Kow	2.29	Log Kow Exp Ref	HANSCH,C ET AL. (1995)
Log Kow Exp	2.36		

*EPI WSKOW model*

*LogKow: log octanol-water partition coefficient*

Kp (m3/ug) Mackay-	0.00666	Kp (m3/ug) Koa-	0.000281
Phi Junge-Pankow-	0.194	Phi Mackay-based	0.347
Phi Koa-based	0.022		

*EPI AEROWIN models*

*Kp: particle-gas partition coefficient. Phi: fraction of substance sorbed to atmospheric particulates*

Koc from MCI (L/kg)	31.27	Log Koc from MCI	1.4951
Koc from Kow (L/kg)	101.8	Log Koc from Kow	2.0076

*EPI KOCWIN models*

*Koc: soil adsorption coefficient of organic compounds. Kow: octanol-water partition coefficient. MCI: first order Molecular Connectivity Index*

Level III Fugacity Environmental Partitioning, emission to air, water and soil

	Air	Water	Soil	Sediment
Mass Amount (%)	0.0763	26	73.9	0.0814
Half-Life (hr)	3.32	360	720	3240
Emissions (kg/hr)	1000	1000	1000	0

*EPI Level III Fugacity Model*

Persistence time (hr)	610
Persistence time (days)	25.41667

*EPI Level III Fugacity Model*

Level III Fugacity Environmental Partitioning, emission only to water

	Air	Water	Soil	Sediment
Mass Amount (%)	6.95E-006	99.7	0.0026	0.312
Half-Life (hr)	3.32	360	720	3240
Emissions (kg/hr)	0	1000	0	0

*EPI Level III Fugacity Model*

Persistence time (hr)	343
Persistence time (days)	14.29167

*EPI Level III Fugacity Model*

Sewage Treatment Plant (STP) overall chemical mass balance using 10,000 hr

	Total removal	Biodegradation	Sludge Adsorption	Volatilization
(%)	2.76	0.1	2.66	0

*EPI STPWIN model*

Atmospheric oxidation (25 deg C)

	OH	Ozone
Half-Life (d)	0.1382	0
Half-Life (hr)	1.658	
Overall Rate Const. (OH: E-12 cm <sup>3</sup> /molecule-sec and OZ: E-17 cm <sup>3</sup> /molecule-sec)	77.4198	

*EPI AOPWIN models*

Biodegradation

Biowin1 (linear model) Probability of Rapid	1.2526
Biowin2 (non-linear model) Probability of Rapid	1
Biowin3 Expert Survey Ultimate Biodegradation	2.9033
Biowin3 Expert Survey Ultimate Timeframe	weeks
Biowin4 Expert Survey Primary Biodegradation	4.2944
Biowin4 Exp. Survey Primary Timeframe	hours-days
Biowin5 (MITI linear model) Biodegradation	0.6707
Biowin6 (MITI non-linear model) Biodegradation	0.5691
Biowin7 (Anaerobic Linear) Biodegradation	1.3006

Petroleum Hydrocarbon Biodegradation Half-Life

*EPI BIOWIN models*

*SkinBiowin1 and Biowin2: ≥0.5: "Rapid" <0.5: "Slow"*

*Biowin3 and Biowin4: 5 ~ hours; 4 ~ days; 3 ~ weeks; 2 ~ months; 1 ~ years.*

*Biowin5 and Biowin6: ≥0.5: "Readily", <0.5: "Not readily".*

	Exp	Battery	CASE Ultra	Leadscope	SciQSAR
Not Ready Biodegradability (POS=Not Ready)		INC_OUT	POS_IN	NEG_IN	INC_OUT

*DTU-developed models*

## Bioaccumulation

BCF (L/kg wet-wt)	16.75
Log BCF (L/kg wet-wt)	1.224
Whole Body Primary Biotransformation Fisk, Fish Half-Life	0.007579
BCF Arnot-Gobas (upper trophic) Including Biotransformation	24.39
BCF Arnot-Gobas (upper trophic) Zero Biotransformation	25.38
BAF Arnot-Gobas (upper trophic) Including Biotransformation	24.39
BAF Arnot-Gobas (upper trophic) Zero Biotransformation	26.4

### *EPI BCFBAF models*

*BCF: Bioconcentration factor, BAF: Bioaccumulation factor*

## Aquatic toxicity

	Exp	Battery	Leadscope	SciQSAR
Fathead minnow 96h LC50 (mg/L)	14.09205	5.942004	1.745494	10.13851
Domain		IN	IN	IN
Daphnia magna 48h EC50 (mg/L)	0.00220011	0.7453111	1.328652	0.1619699
Domain		IN	IN	IN
Pseudokirchneriella s. 72h EC50 (mg/L)	24.00017	9.340055	17.46523	1.214883
Domain		IN	IN	IN

### *DTU-developed models*

	Fisk, Fish 96h	Daphnid 48h	Green Algae 96h
LC50 (Fisk, Fish) or EC50 (Daphnid and Algae) for Most Toxic Class (mg/L)	1.39	0.003	14.993
Max. Log Kow for Most Toxic Class	5	5	6.4
Most Toxic Class	Esters, Dithiophosphate s	Esters, Dithiophosphate s	Esters, Dithiophosphate s

### Note

#### *EPI ECOSAR models*

*ECOSAR Classes: Esters;Esters, Dithiophosphates*

### Oral absorption

Lipinski's Rule-of-five score (bioavailability)	0
Absorption from gastrointestinal tract for 1 mg dose (%)	95
Absorption from gastrointestinal tract for 1000 mg dose (%)	50

*Leadscope model on Lipinski's Rule-of-five. Equation from literature on GI abs. Lipinski scores of 0 or 1: The substance may be bioavailable. Lipinski scores of 2, 3 or 4: The substance may not be bioavailable.*

### Skin absorption

Dermal absorption (mg/cm <sup>2</sup> /event)	0.00037
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*EPI DERMWIN model*

### Brain/blood Distribution

Log brain/blood partition coefficient	-0.1886
---------------------------------------	---------

*Equation from literature Partitioning between the two tissues at equilibrium. >1: high, >0 to <1: medium, >-1 to <0, fair, <-1: low.*

### Metabolism

	Exp	Battery	CASE Ultra	Leadscope	SciQSAR
CYP2C9 substrates (Human clinical data)	NEG	NEG_IN	NEG_IN	NEG_IN	POS_IN
CYP2D6 substrates (Human clinical data)	NEG	NEG_IN	NEG_IN	NEG_IN	NEG_IN

*DTU-developed models*

### Acute toxicity in Rodents

	LD50 (mg/kg/d)	Reliability Index
Rat Oral	1600	0.58
Rat Intraperitoneal	120	0.59
Mouse Oral	1100	0.39
Mouse Intraperitoneal	920	0.38
Mouse Intravenous	210	0.76
Mouse Subcutaneous	120	0.57

*ACDLabs models Reliability index: <0.3 = Not reliable prediction quality; 0.3-0.5 = borderline prediction quality; 0.5-0.75 = moderate prediction quality; >0.75 = high prediction quality.*

## MRDD in Humans

	Exp	Battery	CASE Ultra	Leadscope	SciQSAR
MRDD in Humans $\leq$ 2.69 mg/kg-bw/d		NEG_IN	INC_OUT	NEG_IN	NEG_IN

### DTU-developed models

Model based on data on pharmaceuticals. Maximum recommended daily dose in pharmaceutical clinical trials employing primarily oral route of exposure and daily treatments, usually for 3-12 months.

## Irritation and Sensitization

	Exp	Battery	CASE Ultra	Leadscope	SciQSAR
Severe Skin Irritation in Rabbit		NEG_IN	INC_OUT	NEG_IN	NEG_IN
Allergic Contact Dermatitis in Guinea Pig and Human*	N/A	INC_OUT	POS_IN	NEG_IN	INC_OUT
Respiratory Sensitisation in Humans		NEG_OU T	INC_OUT	NEG_IN	INC_OUT

### DTU-developed models

\*Based on commercial training set

	VEGA	ADI
Skin Sensitization (CAESAR)	NEG_Low	0

CAESAR skin sensitization model is version 2.1.6 contained in VEGA command line version 1.1.2 BETA 5 with calculation core version 1.2.4

Prediction: POS = Sensitizer, NEG = Non-sensitizer, SUSP.POS = Suspected sensitizer, POSS.NEG = Possible Non-sensitizer, Exp = experimental value, Good = Good reliability, Mod = Moderate reliability, Low = Low reliability.

### Protein binding by OASIS, alerts in:

- parent only (Thio)Phosphates
- metabolites from skin metabolism simulator only (Thio)Phosphates
- metabolites from auto-oxidation simulator only

### Protein binding by OECD, alerts in:

- parent only No alert found
- metabolites from skin metabolism simulator only No alert found
- metabolites from auto-oxidation simulator only

### Protein binding potency Cys (DRPA 13%), alerts in:

- parent only DPRA less than 9% (DPRA 13%) >> Non-



	Conjugated carboxylic acids and esters (non reactive)
- metabolites from skin metabolism simulator only	DPRA less than 9% (DPRA 13%) >> Alcohols; DPRA less than 9% (DPRA 13%) >> Non-Conjugated carboxylic acids and esters (non reactive)
- metabolites from auto-oxidation simulator only	
Protein binding potency Lys (DRPA 13%), alerts in:	
- parent only	DPRA less than 9% (DPRA 13%) >> Non-Conjugated carboxylic acids and esters (non reactive)
- metabolites from skin metabolism simulator only	DPRA less than 9% (DPRA 13%) >> Alcohols; DPRA less than 9% (DPRA 13%) >> Non-Conjugated carboxylic acids and esters (non reactive); DPRA less than 9% (DPRA 13%) >> Nonionic surfactants
- metabolites from auto-oxidation simulator only	
Keratinocyte gene expression, alerts in:	
- parent only	Not possible to classify according to these rules
- metabolites from skin metabolism simulator only	Not possible to classify according to these rules
- metabolites from auto-oxidation simulator only	
Protein binding potency GSH, alerts in:	
- parent only	Not possible to classify according to these rules (GSH)

#### OECD QSAR Toolbox v.4.1 profilers

Profiler predictions are supporting information to be used together with the relevant QSAR predictions

#### Endocrine and Molecular Endpoints

	Exp	Battery	CASE Ultra	Leadscope	SciQSAR
Estrogen Receptor $\alpha$ Binding, Full training set (Human <i>in vitro</i> )	NEG	NEG_IN	NEG_IN	NEG_IN	POS_IN
Estrogen Receptor $\alpha$ Binding, Balanced Training Set (Human <i>in vitro</i> )	NEG	NEG_IN	NEG_IN	NEG_IN	NEG_IN
Estrogen Receptor $\alpha$ Activation (Human <i>in vitro</i> )		NEG_IN	INC_OUT	NEG_IN	NEG_IN
Estrogen Receptor Activation, CERAPP data ( <i>in vitro</i> )	NEG	N/A	N/A	NEG_IN	N/A
Androgen Receptor Inhibition	NEG	NEG_IN	NEG_IN	NEG_IN	NEG_IN

	Exp	Battery	CASE Ultra	Leadscop e	SciQSAR
<i>(Human in vitro)</i>					
Androgen Receptor Binding, CoMPARA data ( <i>in vitro</i> )	NEG	N/A	N/A	NEG_IN	N/A
Androgen Receptor Inhibition, CoMPARA data ( <i>in vitro</i> )	NEG	N/A	N/A	NEG_IN	N/A
Androgen Receptor Activation, CoMPARA data ( <i>in vitro</i> )	NEG	N/A	N/A	NEG_IN	N/A
Thyroperoxidase (TPO) inhibition QSAR1 (Rat <i>in vitro</i> )	POS	N/A	N/A	NEG_OUT	N/A
Thyroperoxidase (TPO) inhibition QSAR2 (Rat <i>in vitro</i> )	POS	N/A	N/A	NEG_OUT	N/A
Sodium/iodide symporter (NIS), higher sensitivity		N/A	N/A	NEG_IN	N/A
Sodium/iodide symporter (NIS), higher specificity		N/A	N/A	NEG_IN	N/A
Thyroid Receptor $\alpha$ Binding (Human <i>in vitro</i> )					
mg/L			52841.4	4745.745	951.2055
$\mu$ M			159955.8	14365.81	2879.387
Positive for $IC_{50} \leq 10 \mu$ M					
Positive for $IC_{50} \leq 100 \mu$ M					
Domain		OUT	OUT	OUT	OUT
Thyroid Receptor $\beta$ Binding (Human <i>in vitro</i> )					
mg/L			10689.92	270.8846	142.7823
$\mu$ M			32359.38	819.9927	432.2151
Positive for $IC_{50} \leq 10 \mu$ M					
Positive for $IC_{50} \leq 100 \mu$ M					
Domain		OUT	OUT	OUT	OUT
Arylhydrocarbon (AhR) Activation – Rational final model (Human <i>in vitro</i> )					
		N/A	N/A	NEG_IN	N/A
Arylhydrocarbon (AhR) Activation – Random final model (Human <i>in vitro</i> )					
		N/A	N/A	NEG_IN	N/A
Pregnane X Receptor (PXR) Binding (Human <i>in vitro</i> )	N/A	INC_OUT	INC_OUT	POS_OUT	INC_OUT
Pregnane X Receptor (PXR) Binding (Human <i>in vitro</i> ) NEW		N/A	N/A	INC_OUT	N/A
Pregnane X Receptor (PXR) Activation (Human <i>in vitro</i> )	NEG	N/A	N/A	NEG_OUT	N/A
Pregnane X Receptor (PXR) Activation (Rat <i>in vitro</i> )	NEG	N/A	N/A	NEG_IN	N/A
CYP3A4 Induction (Human <i>in vitro</i> )	NEG	N/A	N/A	NEG_IN	N/A
Constitutive Androstane Receptor (CAR) Activation at max. 20 $\mu$ M ( <i>in vitro</i> )		N/A	N/A	NEG_IN	N/A

	Exp	Battery	CASE Ultra	Leadscope	SciQSAR
Constitutive Androstane Receptor (CAR) Activation at max. 50 µM ( <i>in vitro</i> )		N/A	N/A	NEG_IN	N/A
Constitutive Androstane Receptor (CAR) Inhibition at max. 20 µM ( <i>in vitro</i> )		N/A	N/A	NEG_IN	N/A
Constitutive Androstane Receptor (CAR) Inhibition at max. 50 µM ( <i>in vitro</i> )		N/A	N/A	NEG_IN	N/A

*DTU-developed models*

Estrogen Receptor Binding, alerts in:	
- parent only	Non binder, non cyclic structure
- metabolites from <i>in vivo</i> Rat metabolism simulator only	Non binder, non cyclic structure
- metabolites from Rat liver S9 metabolism simulator only	Non binder, non cyclic structure
rtER Expert System - USEPA, alerts in:	
- parent only	No alert found
- metabolites from <i>in vivo</i> Rat metabolism simulator only	No alert found
- metabolites from Rat liver S9 metabolism simulator only	No alert found

*OECD QSAR Toolbox v.4.2 profilers*

*Profiler predictions are supporting information to be used together with the relevant QSAR predictions*

#### Developmental Toxicity

	Battery	CASE Ultra	Leadscope	SciQSAR
Teratogenic Potential in Humans	NEG_IN	NEG_IN	NEG_IN	NEG_IN

*DTU-developed models based on commercial training set*

#### Genotoxicity - Structural Alerts for DNA Reactivity

	Battery	CASE Ultra	Leadscope	SciQSAR
Ashby Structural Alerts	POS_IN	POS_IN	POS_IN	POS_IN

*DTU-developed models based on commercial training set*

DNA binding by OASIS, alerts in:	
- parent only	Alkylphosphates, Alkylthiophosphates and Alkylphosphonates
DNA binding by OECD, alerts in:	
- parent only	No alert found

*OECD QSAR Toolbox v.4.2 profilers*

*Profiler predictions are supporting information to be used together with the relevant QSAR predictions*

*In vitro* Genotoxicity - Bacterial Reverse Mutation Test (Ames test)

	Exp	Battery	CASE Ultra	Leadscope	SciQSAR
Ames test in <i>S. typhimurium</i> ( <i>in vitro</i> )	NEG	NEG_IN	NEG_IN	NEG_IN	NEG_OUT
*Direct Acting Mutagens (without S9)	N/A	POS_OUT	INC_OUT	POS_IN	NEG_OUT
*Base-Pair Ames Mutagens	N/A	NEG_IN	INC_OUT	NEG_IN	NEG_IN
*Frameshift Ames Mutagens	N/A	NEG_OUT	INC_OUT	INC_OUT	NEG_IN
*Potent Ames Mutagens, Reversions $\geq$ 10 Times Controls	N/A	INC_OUT	INC_OUT	INC_OUT	INC_OUT

*DTU-developed models*

\* The four models (Direct Acting mutagens (without S9), Base-Pair Ames Mutagens, Frameshift Ames Mutagens, Potent Ames Mutagens) should not be used to determine if substances are Ames mutagens, but can be used for indication of mechanism or potency for cases where the main Ames model (Ames test in *S. typhimurium* (*in vitro*)) is POS\_IN.

	VEGA	Mut. / Non-mut. scores	Used models
Mutagenicity consensus	NEG	0 / 1	3

*Mutagenicity (Ames) consensus model version 1.0.2 contained in VEGA version 1.1.4 with calculation core version 1.2.4*

*Prediction: POS = Mutagenic, NEG = Non-mutagenic.*

VEGA

ISS	CAESAR	SarPy	KNN
NEG_Exp	NEG_Exp	NEG_Exp	NEG_Low

*Four individual models in mutagenicity consensus model version 1.0.2 contained in VEGA version 1.1.4 with calculation core version 1.2.4*

*Prediction: POS = Mutagenic, NEG = Non-mutagenic, SUSP.POS = Suspected mutagenic, POSS.NEG = Possible Non-mutagenic, Exp = experimental value, Good = Good reliability, Mod = Moderate reliability, Low = Low reliability.*

DNA alerts for AMES by OASIS, alerts in:

- parent only No alert found

*In vitro* mutagenicity (Ames test) alerts by ISS, alerts in:

- parent only No alert found

*OECD QSAR Toolbox v.4.2 profilers*

*Profiler predictions are supporting information to be used together with the relevant QSAR predictions*

## Other *in vitro* Genotoxicity Endpoints

	Exp	Battery	CASE Ultra	Leadscope	SciQSAR
Chromosome Aberrations in Chinese Hamster Ovary (CHO) Cells*	N/A	POS_IN	POS_IN	NEG_IN	POS_IN
Chromosome Aberrations in Chinese Hamster Lung (CHL) Cells	POS	POS_IN	POS_IN	NEG_OUT	POS_IN
Mutations in Thymidine Kinase Locus in Mouse Lymphoma Cells		NEG_OUT	INC_OUT	NEG_IN	INC_OUT
Mutations in HGPRT Locus in Chinese Hamster Ovary (CHO) Cells		INC_OUT	INC_OUT	NEG_IN	POS_IN
Unscheduled DNA Synthesis (UDS) in Rat Hepatocytes		NEG_OUT	INC_OUT	INC_OUT	NEG_IN
Syrian Hamster Embryo (SHE) Cell Transformation		INC_OUT	INC_OUT	NEG_IN	POS_IN

*DTU-developed models*

*\*Based on commercial training set*

*HGPRT: Hypoxanthine-guanine phosphoribosyltransferase*

DNA alerts for CA and MNT by OASIS, alerts in:

- parent only Alkylphosphates, Alkylthiophosphates and Alkylphosphonates

Protein binding alerts for Chromosomal aberration by OASIS, alerts in:

- parent only No alert found

*OECD QSAR Toolbox v.4.2 profilers*

*CA: Chromosomal aberration, MNT: Micronucleus test*

*Profiler predictions are supporting information to be used together with the relevant QSAR predictions*

## *In vivo* Genotoxicity Endpoints

	Exp	Battery	CASE Ultra	Leadscope	SciQSAR
Sex-Linked Recessive Lethal (SLRL) Test in <i>Drosophila m.</i>	NEG	NEG_IN	NEG_IN	NEG_OUT	NEG_IN
Micronucleus Test in Mouse Erythrocytes	POS	POS_IN	POS_IN	NEG_IN	POS_IN
Dominant Lethal Mutations in Rodents		POS_IN	INC_OUT	POS_IN	POS_IN
Sister Chromatid Exchange in Mouse Bone Marrow Cells		INC_OUT	INC_OUT	NEG_IN	POS_IN
Comet Assay in Mouse		INC_OUT	INC_OUT	NEG_IN	POS_IN

*DTU-developed models*

*In vivo* mutagenicity (Micronucleus) alerts by ISS, alerts in:

- parent only H-acceptor-path3-H-acceptor

*OECD QSAR Toolbox v.4.2 profilers*

*Profiler predictions are supporting information to be used together with the relevant QSAR predictions*

#### Carcinogenicity

	E Ultra	Leadscope
FDA RCA Cancer Male Rat	NEG_IN	NEG_IN
FDA RCA Cancer Female Rat	NEG_IN	NEG_IN
FDA RCA Cancer Rat	NEG_IN	NEG_IN
FDA RCA Cancer Male Mouse	INC_IN	NEG_IN
FDA RCA Cancer Female Mouse	NEG_IN	NEG_IN
FDA RCA Cancer Mouse	NEG_IN	NEG_IN
FDA RCA Cancer Rodent	NEG_IN	NEG_IN

*Commercial models from CASE Ultra and Leadscope*

*FDA RCA: Data from US Food and Drug Administration as part of Research Cooperation Agreement*

Carcinogenicity (genotox and nongenotox) alerts by ISS, alerts in:

- parent only No alert found

Oncologic Primary Classification, alerts in:

- parent only Organophosphorus Type Compounds

*OECD QSAR Toolbox v.4.2 profilers*

*Profiler predictions are supporting information to be used together with the relevant QSAR predictions*

	Exp	Battery	CASE Ultra	Leadscope	SciQSA R
Liver Specific Cancer in Rat or Mouse		INC_OUT	INC_OUT	NEG_IN	POS_IN

*DTU-developed models*

#### Abbreviations

INC: inconclusive. A definite call within the defined applicability domain could not be made.

NEG: negative

POS: positive

IN: inside applicability domain

OUT: outside applicability domain

Exp: Experimental values, from EpiSuite experimental databases or DK DTU QSAR models training sets.

N/A: Not applicable, either because training set data cannot be released for commercial or proprietary models / training sets, or because the model was not developed in a given QSAR software (i.e. a given prediction is not available as the model version does not exist).

#### Important notes

This is an automatically generated report from the Danish (Q)SAR Database, <http://qsar.food.dtu.dk>.

For predictions from CASE Ultra, Leadscope, SciQSAR as well as the Acute toxicity in rodent from ACDLabs information on the software versions can be found in the QMRFs. For the other predicted properties the software versions are:

EPI MPBPWIN v1.43  
EPI HENRYWIN v3.20  
EPI WSKOW v1.42  
EPI WATERNT v1.01  
EPI KOAWIN v1.10  
EPI AEROWIN v1.00  
EPI KOCWIN v2.00  
EPI Level III Fugacity Model (EPI Suite v4.11)  
EPI STPWIN (EPI Suite v4.11)  
EPI AOPWIN v1.92  
EPI BIOWIN v4.10  
EPI BCFBAF v3.01  
EPI ECOSAR v1.11  
EPI DERMWIN v2.02  
ACD/ ToxSuite 2.95.1 Ionization\pKa  
ACD/ ToxSuite 2.95.1 Ionization\LogD  
ACD/ ToxSuite 2.95.1

It is recommended to run the latest version of the EPI Suite Programs in preference of the predictions given in this document when these endpoints are of importance and new versions have been released from the United States Environmental Protection Agency in comparisons. EPI Suite can be downloaded from the US EPA homepage: <http://www.epa.gov/oppt/exposure/pubs/episuitedl.htm>

For further information on the applied systems, see the following homepages:

Case Ultra: <http://www.multicase.com/case-ultra>

Leadscope: <http://www.leadscope.com/>

SciQSAR: <http://lhasa-llc.com/>

ToxSuite: <http://www.acdlabs.com/>

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All access to the database should happen through the provided client-side software and without any use of automated workflow or scripting.

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